Mathematical Modelling in Physiology: Biomedical applications Moscow, 21-24.03.2016

Mathematical modelling of immuno-physiological processes in virus infections:

a question driven data-based approach

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The Immune System





Nucleus

Cytoplasm

2)





- Sources of heterogeneity:
- Physical compartments (lymph node, spleen, blood, etc.)
- Distinct cell populations (e.g. lymphocytes, Macrophages)
- *Heterogeneity w.r.t. the expression of specific markers* (CCR5) or fluorescent labels (CFSE, BrdU)

The fundamental challenge to applied mathematicians is to understand immunology in new ways by using models, computational techniques & algorithms

...

1) 2)

3)

4)

Challenges in understanding the IS

"...The many immunological observations and results from in vitro or in vivo experiments vary and their interpretations differ enormously. A major problem is that within a normal distribution of biological phenomena that are measurable with many methods virtually anything can be shown or is possible.." *R. M. Zinkernagel. Immunity Against Infections & Vaccines: Credo 2004. Scand J Immunol. 2004, 60: 9–13*



- Infectious disease model: key components
- Antiviral immune response: <u>kinetic regulation</u>
- Optimal model structure: <u>information-theoretic criteria</u>
- Cell growth analysis: <u>asymmetric division</u>

Dynamic patterns of infectious diseases





- Infectious disease model: <u>key components</u>
- Antiviral immune response: <u>kinetic regulation</u>
- Optimal model structure: <u>information-</u> <u>theoretic criteria</u>
- Cell growth analysis: <u>asymmetric division</u>

The "numbers game" perspective: virus & host factors in the outcome of infection



The clinician's perspective:

- Cytopathicity of virus
- Latency
- Persistence
- <u>Replication rate</u>
- Tropism
- Immunopathology
- Health condition of the infected individual

The mathematical view:

$$\begin{aligned} \frac{d}{dt}V(t) &= \left(\beta - \gamma \cdot F(t)\right) \cdot V(t) \\ \frac{d}{dt}F(t) &= \rho \cdot C(t) - \eta \cdot \gamma \cdot F(t) \cdot V(t) - \mu_f \cdot F(t) \\ \frac{d}{dt}C(t) &= \xi(m) \cdot \alpha \cdot V(t - \tau) \cdot F(t - \tau) - \mu_f \cdot \left(C - C^*\right) \\ \frac{d}{dt}m(t) &= \sigma \cdot V(t) - \mu_m \cdot m(t) \end{aligned}$$

 $V(t_0) = V_0, \quad F(t_0) = F_0, \quad C(t_0) = C_0, \quad m(t_0) = m_0,$ $V(t) = 0, \quad F(t) = F_0 \quad \text{при } t \in [t_0 - \tau, t_0)$

Modelling in immunology: Experimental and Mathematical Approaches

Nobel Prize Laureate Rolf M. Zinkernagel



Academician Guri I. Marchuk

"The outcome of infection results from the *´ numbers games*' between infectious agent and the immune system."

Replication rate in virus persistence: pro & contra



- <u>Experimental studies</u> with LCMV infection in mice: a faster speed of virus replication is an advantage for a virus in overcoming the immune system control and establishing the persistent infection – the tolerance by exhaustion (Moskophidis et al., *Nature* (1993) 362: 758-761)
- <u>Theoretical study of virus infection model</u>: a slow virus replication favors the long-term persistence (Marchuk and Belykh, 1980)

Mathematical immunology and the nuclear chain reaction



George Irving Bell

4.08.1926-28.05.2000

- Harvard University (Physics) 1947
- Division "T" Los Alamos Scientific Laboratory
 1947
- "Nuclear Reactor Theory" 1970
- Quantitative models in immunology 1970
- Theoretical Biology & Biophysics. Los Alamos NL - 1974
- Humane genome Project 1988





Guri Ivanovich Marchuk 8.06.1925-24.03.2013

- Leningrad University (Mathematics/mechanics) 1949
- Division "B" Nuclear Power Plant Ceneter 1953
- "Numerical Methods in the Analysis of the Nuclear Reactors" – M. 1959
- Mathematical modeling in immunology 1975
- Institute of Numerical Mathematics, Academy of Sciences (1980)

Basic scheme of an infectious disease

G.I. Bell:

Predator-prey equations simulating an immune (1973)

- Population dynamics of Replicating antigens and Antibodies
- Lotka-Volterra-type of ODEs

G.I. Marchuk: Mathematical model of infectious disease (1974)

- System of delay differential equations
- Target organ damage
- Negative feedback of the organ damage on the immune system



Basic model of the infectious disease (1975)

State space variable

- 1. Pathogen population
- 2. Antibodies
- 3. Plasma cells
- 4. Tissue damage

System of Delay-Differential Equations $\frac{d}{dt}V(t) = (\beta - \gamma \cdot F(t)) \cdot V(t)$ $\frac{d}{dt}F(t) = \rho \cdot C(t) - \eta \cdot \gamma \cdot F(t) \cdot V(t) - \mu_f \cdot F(t)$ $\frac{d}{dt}C(t) = \xi(m) \cdot \alpha \cdot V(t - \tau) \cdot F(t - \tau) - \mu_f \cdot (C - C^*)$ $\frac{d}{dt}m(t) = \sigma \cdot V(t) - \mu_m \cdot m(t)$

Initial data

$$V(t_0) = V_0, \quad F(t_0) = F_0, \quad C(t_0) = C_0, \quad m(t_0) = m_0,$$

$$V(t) = 0, \quad F(t) = F_0 \quad \text{for } t \in [t_0 - \tau, t_0]$$







Fundamental modelling issues

- The immune system can be considered as *an ensemble of individual cells*, which differ in a number of physiologically relevant traits: the spatial position (*z*), the receptors' specificity (*s*), the number of specific receptors (*r*), etc. => *Heterogeneity*
- One can describe the immune system by a state vector listing the number of individual cells of each "type" n(t, Z, s, r, ...) depending on a number of continuous as well as discrete variables
- Using conservation principle the equation for the density function n(t, x) with x standing for the vector of traits can be written as

 $\frac{\partial n}{\partial t} + \nabla(\mathbf{v}n) = \mathbf{Birth} - \mathbf{Death} + \mathbf{Migration} + \mathbf{Reactions}, \quad \nabla = \left(\frac{\partial}{\partial x_1}, \frac{\partial}{\partial x_2}, \dots, \frac{\partial}{\partial x_K}\right), \quad \mathbf{v} = \left[v_1, v_2, \dots, v_K\right]^{\mathrm{T}}, \quad v_i = \frac{dx_i}{dt}$

• "Typical" equation for population dynamics:

 $\frac{d}{dt}$ (Population size) = **f**(growth, death, migration,...)

Maximum likelihood parameter estimation

$$\frac{d}{dt}\mathbf{y}(t) = \mathbf{f}(\mathbf{y}(t), \mathbf{y}(t-\tau), \mathbf{p}), \quad t \in [t_0, T], \quad \tau > 0$$
$$\mathbf{y} \in R^{n_{\mathbf{Y}}}, \quad \mathbf{p} \in R^{n_{\mathbf{p}}}$$
$$\mathbf{y}(t) = \varphi(t), t \in [t_0 - \tau, t_0].$$

•Observation data

$$\left\{t_{j},\mathbf{y}_{j}\right\}_{j=1}^{n_{obs}}$$

•Likelihood function

$$\mathbf{y}_{j} \sim \mathcal{N}(\mathbf{y}(t_{j}), \Sigma_{j})$$
$$\mathcal{H}(\mathbf{y}_{j}; \mathbf{p}) = \frac{1}{\sqrt{(2\pi)^{n_{y}} \det \Sigma_{j}}} \exp\left\{-\frac{1}{2}[\mathbf{y}(t_{j}) - \mathbf{y}_{j}]^{\mathrm{T}} \Sigma_{j}^{-1}[\mathbf{y}(t_{j}) - \mathbf{y}_{j}]\right\}$$
$$\mathcal{L}(\mathbf{p}) = \prod_{j=1}^{n_{obs}} \mathcal{H}(\mathbf{y}_{j}; \mathbf{p})$$
$$\mathbf{p}^{*} = \arg \max_{\mathbf{p} \in D} \mathcal{L}(\mathbf{p})$$



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Experimental System: Lymphocytic choriomeningitis virus (LCMV) infection of C57BL/6 mice

- Family: arenaviruses
- **Strains:** *Docile, Traub, WE, Aggressive, Armstrong, Clone* 13
- Host: mice, hamsters
 (Humans: acute hemorrhagic fevers Lassa fever)
- Target cells: primarily infects & replicates in macrophages, lymphocytes
- **Cytopathicity in vivo:** *non-cytopathic RNA virus*
- CTL responses play a dominant role in virus clearance: <u>appear</u> <u>early and high</u>
- Neutralizing antibody response: appear only *late* after infection
- Immunopathology: *spleen, liver, central nervous system, etc.*

LCMV infection of mice: a "Gold Standard" of experimental immunology

Relevant patterns of LCMV infection dynamics

The kinetics of the T cell response falls in between two extremes expansion=>contraction=>





Phenomenology of the LCMV-host interaction

Phenotypes of "virus-host" relationships:

- 1. acute infection followed by LCMV clearance in two weeks
- 2. temporary persistence up to 2 or 3 months
- 3. life-long persistence

Final stages of antiviral CTL responses:

- 1. Creation of the CTL memory
- 2. Almost complete loss of the entire LCMV-specific precursor CTRL repertoire
- 3. Complete exhaustion of precursor CTLs

Persistence:

depends on particular combination of virus & host parameters

Mechanistic view of the LCMV infection fate decision



Time-dependent variables of the DDE model

 $V(t) - \underline{\text{virus}}$ titer in spleen at time t (pfu/ml)

 $E_p(t)$ – number of virus-specific **precursor CTLs** in spleen at time t (cell/ml)

E(t) – number of virus-specific <u>effector CTLs</u> in spleen at time t (*cell/ml*) W(t) – cumulative virus antigen load in spleen at time t



Model calibration

CTL RESPONSES IN LCMV INFECTION



Virus growth rate and CTL expansion

Fig. Clonal expansion of gp-33 specific CTL after i.v. infection with 200 pfu of LCMV-WE versus LCMV-Armstrong



Underwhelming the Immune Response: Effect of Slow Virus Growth on CD8⁺-T-Lymphocyte Responses



FIG. 2. Bell-shaped relationship between initial virus growth rate and peak CTL responses in spleen (A) and blood (B) established using data on CTL expansion in C57BL/6 mice infected with distinct LCMV strains exhibiting different growth rates. The error bars indicate standard errors. Variation in the LCMV growth rate can have either an enhancing or a down-regulating effect on the clonal burst size of virusspecific CTLs. Virus replication above a threshold is required for strong CTL response. LCMV-WE appears to be an "optimally" replicating strain in C57BL/6 mice, as it induces the strongest CTL response.

Gennady Bocharov,^{1,2}†‡ Burkhard Ludewig,³†* Antonio Bertoletti,⁴ Paul Klenerman,⁵ Tobias Junt,³ Philippe Krebs,³‡ Tatyana Luzyanina,⁶ Cristophe Fraser,¹ and Roy M. Anderson¹

Published clinical observations: the effect of the virus growth rate



a)

A reduction in virus growth rate between infected subjects is associated with a spectrum of disease outcomes:

fulminant hepatitis (τ₂~ 2.1 to 6.2 days)
acute resolving hepatitis (τ₂~ 2.3 to 10 days)
chronic progressing hepatitis (τ₂~ 4 to 615 days)

The genomic analysis of immune response in chimpanzees infected with HCV virus showed that HCV **replication below a threshold level** required for activation of genes involved in antigen presentation **results in virus persistence** (Su et al. *PNAS USA* (2002): **99**: 15669-15674)

Farci et al. *Science* (2000) **288**: 339-344 Webster et al. *Hepatology* (2000) **32**: 1117-1124

Outcomes of hepatitis B and C virus infections

Slowly does it...



Virus growth rate

- Viruses that replicate slowly invoke a weak adaptive immune response
- A *bell-shaped* relationship between the virus growth rate and the peak T cell response
- Speed of viral replication represents an important *'kinetic' mechanism* influencing the pathogenesis and duration of virus persistence within the human host

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IMMUNE EVASION

Slowly does it... A new study published in the Journal

of Virology shows that viruses that replicate slowly invoke a weak adaptive immune response, specifically a weak cytotoxic T-lymphocyte response, which could contribute to virus persistence and chronic disease.

Two facets of the CTL response can affect virus clearance. Specific CTL clones are amplified in response to antigen stimulation the magnitude of amplification of CTLs increases with increasing antigen concentrations. Viruses that replicate rapidly produce large amounts of antigen, which can overwhelm the specific CTL response. This physical deletion of CTLs known as exhaustion — results in virus persistence in the replication rate of the virus.

Using lymphocytic choriomeningitis virus (LCMV) infection of mice as a model, the amplification of CTLs in response to LCMV strains that have different replication rates was assessed. A bell-shaped response was found: both slow and fast replicating virus strains produced weaker CD8+ T-cell responses compared with a strain that had an intermediate replication rate.

What about hepatitis C virus, which replicates more slowly than LCMV? Available data sets were analysed, and slower virus replication correlated with virus persistence. For hepatitis B virus, one welldocumented study of virus kinetics and CTL response has been analysed. A predator-prey model was constructed by Bocharov et al. - with

as prey — and calibrated using this available HBV data set. The model was used to predict

the effect of changes in virus replication kinetics on the CTL response. Reducing the virus replication rate mented for other important viruses. led to a weaker CTL response, which could result in virus persistence. Certain individuals - 'highresponders' - have a more efficient CTL response, presumably through genetic variation. Even with a simulated high-responder, a slowly replicating virus strain elicited only a downregulate virus replication could weak and transient CTL response. This model predicts that the transition from acute to chronic HBV infection could result from a decrease

The mathematical model used is

reductionist and cannot take into

the CTLs as predators, and the virus account every aspect of the complex interactions between the virus and the immune system. However, models are useful for predicting and planning experimental work, and this modelling approach could be implesuch as HIV or cytomegalovirus. This report clearly indicates that the kinetics of virus replication could be important for the outcome of the infection. Slow viruses could sneak past immune surveillance and establish persistence, and therapies that also result in virus persistence and chronic disease

HIGHLIGHTS

Susan Jone

 References and links
 ORIGINAL RESEARCH PAPER Bocharov, G. et a Underwhelming the immune response: effect of slow virus growth on CD81-T-lymphocyte

responses. J. Virol. 78, 2247-2254 (2004)



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Integration of mathematical modelling into the immunological mainstream

William E. Paul (June 12, 1936 – September 18, 2015)

Feedback regulation of proliferation vs. differentiation rates explains the dependence of CD4 T-cell expansion on precursor number

Gennady Bocharov^{a,1,2}, Juan Quiel^b, Tatyana Luzyanina^c, Hagit Alon^d, Egor Chiglintsev^e, Valery Chereshnev^f, Martin Meier-Schellersheim^b, William E. Paul^{b,2}, and Zvi Grossman^{b,d,1,2}

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Contributed by William E. Paul, January 7, 2011 (sent for review November 16, 2010)



Antigen-stimulated CD4 T-cell expansion is inversely and log-linearly related to precursor number

Juan Quiel^a, Stephane Caucheteux^a, Arian Laurence^b, Nevil J. Singh^c, Gennady Bocharov^d, Shlomo Z. Ben-Sasson^a, Zvi Grossman^a, and William E. Paul^{a,1}

Laboratories of *Immunology and ^cCellular and Molecular Immunology, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, MD 2009;2, ^{bM}Nolecular Immunology and Inflammation Branch, National Institute of Arthritis and Musculoskeletal and Skin Diseases, National Institutes of Health, Bethesda, MD 2009;2 and ⁴Institute of Numerical Mathematics, Russian Academy of Sciences, Moscow 119333, Russia

Contributed by William E. Paul, January 7, 2011 (sent for review November 11, 2010)

Thus, the immune system offers challenges sufficient to test the growing power of mathematical attack on a biological problem. It is to the quantitative prediction of the outcome of given perturbations in the immune system that we envisage our mathematical/ modeling colleagues will apply themselves. There is a richness of opportunities and a myriad of challenges. Good luck!!!!! Math. Model. Nat. Phenom.

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Topic

- Infectious disease model: <u>key components</u>
- Antiviral immune response: kinetic regulation
- Optimal model structure: <u>information-</u> <u>theoretic criteria</u>
- Cell growth analysis: <u>asymmetric division</u>

- "Typical" equation for population dynamics: $\frac{\Delta N}{\Delta t} = \mathbf{f}(growth, death, migration,...)$
- **General problem:** various functional forms can be suggested for the same process...
 - *Which model is better:* that having the fewer parameters or that based more closely on the biology of the system?
 - Is it possible to distinguish between the important and the unimportant assumptions in a model?

Virus - CTL dynamics: typical kinetics



Hierarchy of models for LCMV-CTL data

	5 parameters	$\frac{d}{dt}V(t) = \beta \cdot V(t) \cdot (1 - V(t) / K) - \gamma \cdot V(t) \cdot E(t)$ $\frac{d}{dt}E(t) = b_1 \cdot V(t) \cdot E(t) - \alpha_E \cdot E(t)$	Predator-Prey
xity	6 parameters	$\frac{\frac{d}{dt}V(t) = \beta \cdot V(t) \cdot (1 - V(t) / K) - \gamma \cdot V(t) \cdot E(t)}{\frac{d}{dt}E(t) = b_2 \frac{V(t)}{\theta_{sat} + V(t)}E(t) - \alpha_E \cdot E(t)}$	PP+Holling type II response
Comple	7 parameters	$\frac{\frac{d}{dt}V(t) = \beta \cdot V(t) \cdot (1 - V(t) / K) - \gamma \cdot V(t) \cdot E(t)}{\frac{d}{dt}E(t) = b_2 \frac{V(t - \tau)}{\theta_{sat} + V(t)} E(t - \tau) - \alpha_E \cdot E(t)}$	+ time lag in the CTL division
	8 parameters	$\frac{d}{dt}V(t) = \beta \cdot V(t) \cdot (1 - V(t)/K) - \gamma \cdot V(t) \cdot E(t)$ $\frac{d}{dt}E(t) = b_2 \frac{V(t - \tau)}{\theta_{sat} + V(t)}E(t - \tau) - \alpha_E \cdot E(t) + T^*$	+ CTL homeostasis
	9 parameters	$\frac{d}{dt}V(t) = \beta \cdot V(t) \cdot \left(1 - \frac{V(t)}{K}\right) - \gamma \cdot V(t) \cdot E(t)$ $\frac{d}{dt}E(t) = b_1 \cdot V(t) \cdot E(t) - \alpha_E \cdot E(t) + T^* - r_m \cdot E(t)$	+ CTL memory population
		$\frac{a}{dt}E_m(t) = r_m \cdot E(t) - \alpha_m \cdot E_m(t)$	

Model selection: accuracy & parsimony

Given a set of related models – how to rank them by giving each a score?

• the goodness of fit – the size of the minimized least squares

(or maximized likelihood) function

• the principle parsimony – a proper balance between under-fitting and over-fitting

General principle:

A model should be "<u>as simple as possible yet as complex as necessary"</u> (Denis Noble et al. 2005) with respect to the included variables, model structure and the number of parameters for adequate representation of the data



Criteria for discriminating between models – model selection

• Kullback-Leibler information distance between models f and g is defined as the multidimensional integral:

$$I(f,g) = \int f(x) \log\left(\frac{f(x)}{g(x|\theta)}\right) dx$$

•Akaike (1973) found a rigorous way to estimate the Kullback-Leibler directed distance of the candidate model g to the truth f model based on the empirical log-likelihood function $\mathcal{L}(\mathbf{p})$ at its maximum point $\mathcal{L}_* = \mathcal{L}(\mathbf{p}_*)$

•The *Akaike's Information Criterion* makes use of an estimate of the *expected*, *relative distance* between the fitted model and the unknown true mechanism that actually generated the *observed* data (n_{obs})

$$\mu_{AIC} = n_{obs} \cdot \ln(\mathcal{L}(\mathbf{p}^*)) + 2(n_{\mathbf{p}} + 1)$$

•The Kolomogorov complexity – Minimum Description Length principle by Jorma Rissanen "Modelling by the Shortest Data Description" (1978)

$$MDL = -\ln(\mathcal{L}(\mathbf{p}^*)) + 0.5 \cdot n_{\mathbf{p}} \cdot \ln(n_{obs}/(2\pi)) + \ln \int_{\Omega} \sqrt{\det(\mathbf{I}(\mathbf{p}))} d\mathbf{p}$$

Model Ranking

$$\frac{d}{dt}V(t) = \beta \cdot V(t) \cdot (1 - V(t)/K) - \gamma \cdot V(t) \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_1 \cdot V(t) \cdot E(t) - \alpha_E \cdot E(t)$$

$$\frac{d}{dt}V(t) = \beta \cdot V(t) \cdot (1 - V(t)/K) - \gamma \cdot V(t) \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_2 \frac{V(t)}{\theta_{aat} + V(t)}E(t) - \alpha_E \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_2 \frac{V(t - \tau)}{\theta_{aat} + V(t)}E(t - \tau) - \alpha_E \cdot E(t)$$

$$\frac{d}{dt}E(t) = \beta \cdot V(t) \cdot (1 - V(t)/K) - \gamma \cdot V(t) \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_2 \frac{V(t - \tau)}{\theta_{aat} + V(t)}E(t - \tau) - \alpha_E \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_2 \frac{V(t - \tau)}{\theta_{aat} + V(t)}E(t - \tau) - \alpha_E \cdot E(t) + T^*$$

$$\frac{d}{dt}V(t) = \beta \cdot V(t) \cdot (1 - V(t)/K) - \gamma \cdot V(t) \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_2 \frac{V(t - \tau)}{\theta_{aat} + V(t)}E(t - \tau) - \alpha_E \cdot E(t) + T^*$$

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$$\frac{d}{dt}E(t) = b_1 \cdot V(t) \cdot (1 - V(t)/K) - \gamma \cdot V(t) \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_1 \cdot V(t) \cdot E(t) - \alpha_E \cdot E(t) + T^* - r_m \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_1 \cdot V(t) \cdot E(t) - \alpha_E \cdot E(t) + T^* - r_m \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_1 \cdot V(t) \cdot E(t) - \alpha_E \cdot E(t) + T^* - r_m \cdot E(t)$$

$$\frac{d}{dt}E(t) = r_m \cdot E(t) - \alpha_m \cdot E_m(t)$$

$$\frac{d}{dt}E(t) = r_m \cdot E(t) - \alpha_m \cdot E_m(t)$$

$$\frac{d}{dt}E(t) = \frac{d}{dt}E(t) = \frac{d}{dt}E(t) = \frac{d}{dt}E(t) - \frac{d}{dt}E(t) = \frac{d}{dt}E(t) - \frac{d}{dt}E(t) = \frac{d}{dt}E(t) - \frac{d}{dt$$

Uncertainty in parameter estimates: 95% Confidence Intervals

Parameter	Model 1	Model 2
[95%CI]		
Virus growth rate	4.61∈	4.51∈
constant	[3.7, 6.5]	[4.2, 4.8]
CTL activation	9.2x10 ⁻⁷	1.42∈
rate constant	[6.8 •10 ⁻⁷ ,1.4•10 ⁻⁶]	[1.36, 1.48]
CTL death rate	0.093∈	0.2∈
constant	$[10^{-10}, 0.2]$	[0.15, 0.26]

The 'best' model for the given data

$$\beta = 4.51; \quad K = 4.69 \cdot 10^{6}; \quad \gamma = 8 \cdot 10^{-5}$$

$$b_{2} = 1.42; \quad \theta_{sat} = 3.23 \cdot 10^{-176} \approx 0; \quad \alpha_{E} = 0.2$$

$$\frac{d}{dt}V(t) = \beta \cdot V(t) \cdot (1 - V(t)/K) - \gamma \cdot V(t) \cdot E(t)$$

$$\frac{d}{dt}E(t) = b_{2} \frac{V(t)}{\theta_{sat} + V(t)}E(t) - \alpha_{E} \cdot E(t)$$



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Computational approaches to parameter estimation and model selection in immunology

C.T.H. Baker^{a,b,*}, G.A. Bocharov^{a,b,c}, J.M. Ford^d, P.M. Lumb^b, S.J. Norton^b, C.A.H. Paul^a, T. Junt^e, P. Krebs^e, B. Ludewig^{e, f}

$$\frac{d}{dt}E(t) = b_2 \cdot E(t) \cdot H(V - V^*) - \alpha_E \cdot E(t)$$

where $H(\cdot)$ is the Heaviside's step function with

$$V^*$$
 assuming the value ~ $heta_{
m sat}$



Topic

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CFSE Proliferation Assay: analysis of antigen specific and non-specific T cell proliferation.



From: Quah et al., Nat Protoc.2007;2:2049



http://www.biochemmack.ru/product/citometry/cytomet r/CellLabQuantaSC/.

- Major assumption: cell division is symmetric, i.e. the label is halved in the two daughter cells
- Cell biology axiom: a random and uneven partition of mass between the sister cells

R. Sennerstam, partition of (mass) protein to sister cell pairs at mitosis: a re-evaluation. J Cell Sci 1988, 90:301

• Does this matter?



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Low asymmetry of the division: 46% vs 54%









Higher asymmetry of division: 42% vs 58%









Ssoftware for CFSE histogram decomposition



FlowJo is a product of Tree Star, Inc. www.flowjo.com 1-541-201-0022 flowjo@treestar.com TN.Proliferation.20100424

Raw data



Problem: an interpretation of flow cytometry data



6



Time (days)





Proliferation of retrogenic CD8+ T cells



Time (hours)

Division- and CFSE-label structured mathematical model



Log CFSE (UI)

Delay-hPDE model with asymmetric division

J. Math. Biol. (2014) 69:1547–1583 DOI 10.1007/s00285-013-0741-z

Mathematical Biology

$$n_i(t,x) = n_i^r(t,x) + n_i^c(t,x), \quad i = 0,1...,i_p - 1,$$

 $n(t x) = n^{r}(t x)$

Mathematical models for CFSE labelled lymphocyte dynamics: asymmetry and time-lag in division

Tatyana Luzyanina · Jovana Cupovic · Burkhard Ludewig · Gennady Bocharov

$$\begin{aligned} \frac{\partial}{\partial t} n_0^r(t, x) &= n_{i_r}(t, x) \\ \frac{\partial}{\partial t} n_0^r(t, x) - k \frac{\partial}{\partial x} \left(x n_0^r(t, x) \right) &= -(\alpha_0 + \beta_0) n_0^r(t, x), \\ \frac{\partial}{\partial t} n_i^r(t, x) - k \frac{\partial}{\partial x} \left(x n_i^r(t, x) \right) &= -(\alpha_i + \beta_i) n_i^r(t, x) + \\ &+ \alpha_{i-1} e^{k\tau_{i-1}} \left(\frac{1}{m_1} n_{i-1}^r(t - \tau_{i-1}, e^{k\tau_{i-1}} \frac{x}{m_1}) + \frac{1}{1 - m_1} n_{i-1}^r(t - \tau_{i-1}, e^{k\tau_{i-1}} \frac{x}{1 - m_2}) \right), i = 1, 2, ..., i_r, \\ \frac{\partial}{\partial t} n_i^c(t, x) - k \frac{\partial}{\partial x} \left(x n_i^c(t, x) \right) &= \alpha_i \left(n_i^r(t, x) - e^{k\tau_i} n_i^r(t - \tau_i, e^{k\tau_i} x) \right), i = 0, 1, ..., i_r - 1, \\ \frac{\partial}{\partial t} n_i^c(s, x) &= 0, s \in [-\tau_0, 0); n_0^r(0, x) = n^0(x); \\ n_i^r(s, x) &= 0, s \in [-\tau_i, 0], i = 1, 2, ..., i_r; \\ n_i^c(s, x) &= 0, s \in [-\tau_i, 0], i = 0, 1, ..., i_r - 1; \\ n_i^r(t, x_{\max}) &= n_i^c(t, x_{\max}) = 0, t \ge 0; \\ v(0) n_i^r(t, 0) &= v(0) n_i^c(t, x) = 0, t \ge 0; \end{aligned}$$

Label- and division- structured delay model for an asymmetric cell division



Label- and division- structured symmetric cell division



Generations overlap



J. Math. Biol. (2014) 69:1547–1583 DOI 10.1007/s00285-013-0741-z

Mathematical Biology

Mathematical models for CFSE labelled lymphocyte dynamics: asymmetry and time-lag in division

Tatyana Luzyanina $\,\cdot\,$ Jovana Cupovic $\,\cdot\,$ Burkhard Ludewig $\,\cdot\,$ Gennady Bocharov



MINI REVIEW ARTICLE published: 02 September 2013 doi: 10.3389/fimmu.2013.00264



Asymmetry of cell division in CFSE-based lymphocyte proliferation analysis

Gennady Bocharov¹*, Tatyana Luzyanina², Jovana Cupovic³ and Burkhard Ludewig³

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² Institute of Mathematical Problems in Biology, Russian Academy of Sciences, Pushchino, Russia

³ Institute of Immunobiology, Kantonal Hospital St. Gallen, St. Gallen, Switzerland

Embedding the cell population dynamics models into the spatial contexts of the lymphoid organs

Mesoscopic imaging



EYFF

B220

Burkhard Ludewig et al.

DOI: 10.1002/eji.201242508

Eur. J. Immunol. 2012. 42: 3116-3125

Immunology

A global "imaging" view on systems approaches in immunology

"Space" is the current frontier in Systems Immunology

Analyses of the dynamic cell behavior to predict the immune function in health and disease

Technologies to observe and understand cells in motion (Tang et al., 2013, EJI):

• Data capturing and processing: new computational tools to deal with the **vast amount** of data generated by imaging

<u>Capturing the structure-function relationship in a computationally efficient manner is</u> <u>the key to successful systems immunology</u>

 Requires anatomically based mathematical models integrating the interaction processes across multiple scales (multi-physics):



(From: Kumar et al, 2008 EJI)

- Tissue properties
- Blood and lymph flow patterns
- Fluid-tissue interactions
- Systemic cell re-circulation
- Within organs cell migration



3D modelling of the lymph node structure

Math. Model. Nat. Phenom. Vol. 6, No. 7, 2011, pp. 13-26 DOI: 10.1051/mmnp/20116702

Reaction-Diffusion Modelling of Interferon Distribution in Secondary Lymphoid Organs

G. Bocharov^{1*†}, A. Danilov^{1*†}, Yu. Vassilevski¹, G.I. Marchuk¹, V.A. Chereshnev² and B. Ludewig³ ISSN 0012-4966, Doklady Biological Sciences, 2011, Vol. 439, pp. 194–196. © Pleiades Publishing, Ltd., 2011. Original Russian Teer & G.A. Bocharov, A.A. Danilov, Yu.V. Vassilevski, G.I. Marchuk, V.A. Chereshnev, B. Ludewig, 2011, published in Doklady Akademii Nauk, 2011, Vol. 439, No. 3, pp. 413–415.

PHYSIOLOGY =

Simulation of the Interferon-Mediated Protective Field in Lymphoid Organs with Their Spatial and Functional Organization Taken into Consideration

> G. A. Bocharov^a, A. A. Danilov^a, Yu. V. Vassilevski^a, Academician G. I. Marchuk^a, Academician V. A. Chereshnev^b, and B. Ludewig^c

Computation 2015, 3, 222-234; doi:10.3390/computation3020222

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Communication

Computational Approach to 3D Modeling of the Lymph Node Geometry

Alexey Kislitsyn ^{1,†}, Rostislav Savinkov ^{1,†}, Mario Novkovic ^{2,†}, Lucas Onder ² and Gennady Bocharov ^{3,‡}

Submitted:

Data-driven modelling of the FRC network for studying the fluid flow in the conduit system

Rostislav Savinkov^{a,b}, Alexey Kislitsyn^{a,b}, Daniel J. Watson^c, Raoul van Loon^c, Igor Sazonov^c, Mario Novkovic^d, Lucas Onder^d, Gennady Bocharov^{a,*}

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 ^cCollege of Engineering, Swansea University, Swansea, Wales, U.K.
 ^dInstitute of Immunobiology, Kantonsspital St. Gallen, St. Gallen, Switzerland

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- Mathematics Department, The University of Manchester: *Christopher T.H. Baker*
- Mathematics Department, The University of Chester: *Neville J. Ford*
- Institute of Mathematical Problems in Biology, RAS: *Tatyana Luzyanina*
- Children's Hospital, Freiburg University: *Stephan Ehl*
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