

# A hybrid multiscale computational framework to personalize and optimize cancer therapy

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# Context

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Important progress has been made in cancer therapy over the past 10 years

- a wider range of therapeutic options
- more specific molecules that reduce side effects
- more individualized therapies

However, tumour development becomes more complex with therapies

- resistance to treatment
- selection of the most aggressive phenotypes

## **The new objectives**

- rationalize the use of the therapies
- combine the therapies to enhance the effects
- adapt the therapies with the evolution of the tumour and patient states

# Why the need for a virtual tumour ?

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**Provide an integrated vision of the phenomenon**

**Gives the ability to make virtual experiments**

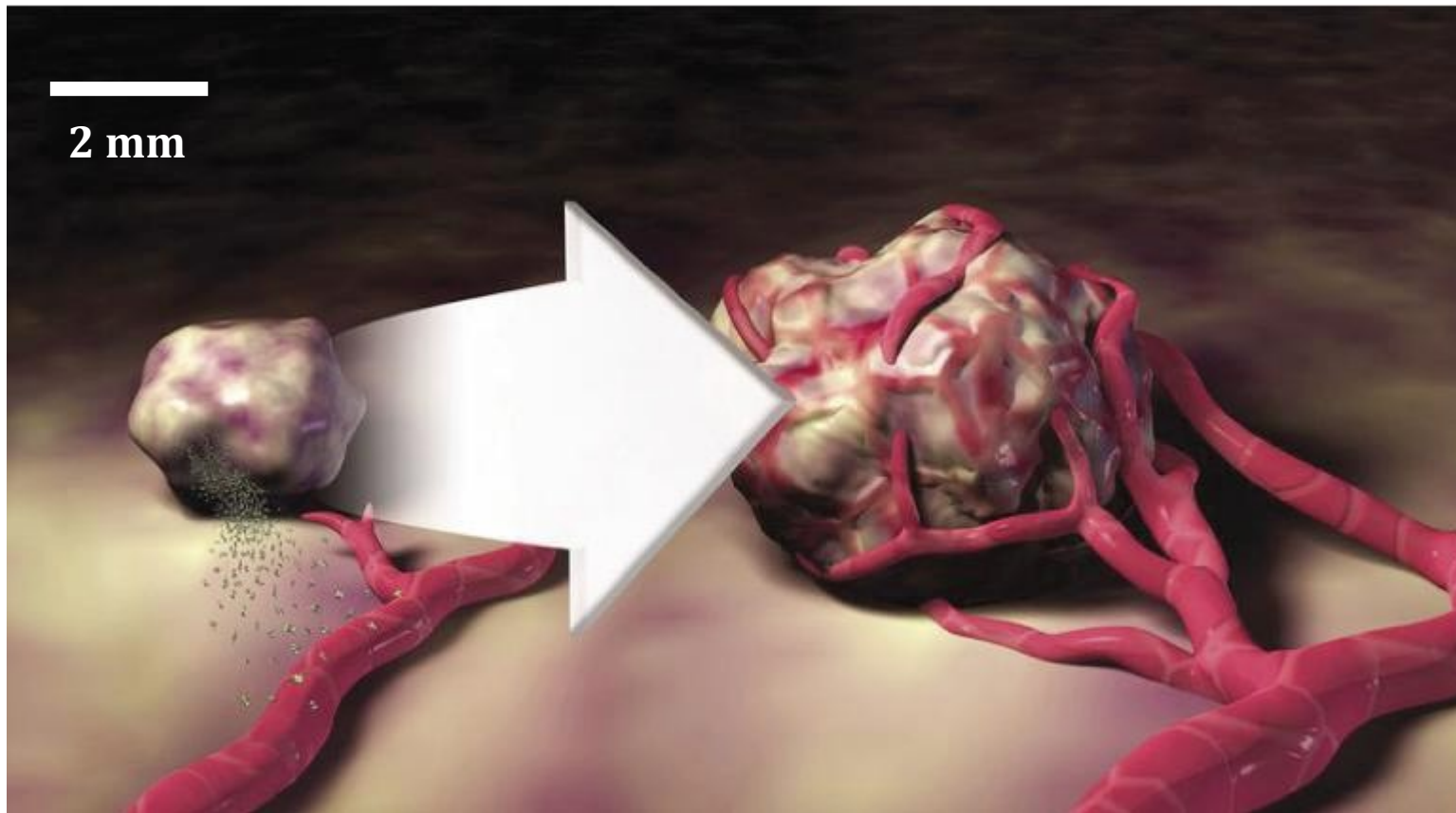
- to test hypotheses
- to better understand the system
- to predict the system's reaction
- to optimize the solutions

# Development of a solid tumour

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**Avascular growth**

**Vascular growth**



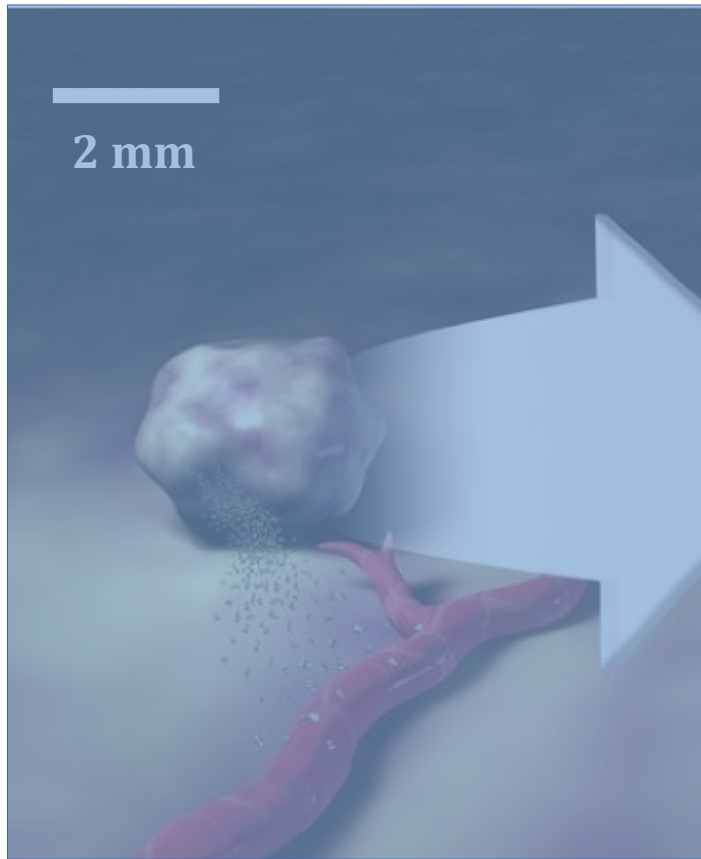
hypoxia induces VEGF release

VEGF induces angiogenesis

# Development of a solid tumour

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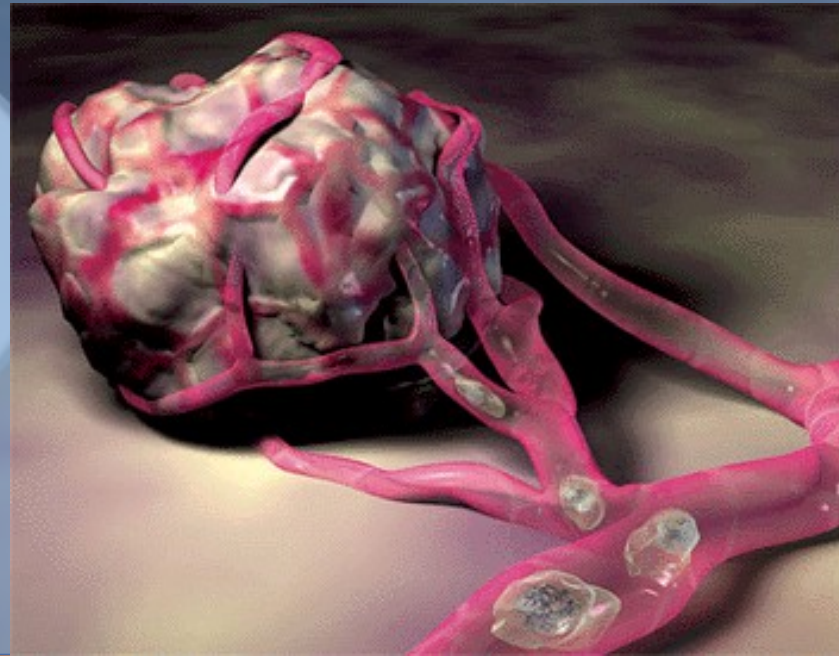
## Avascular growth



hypoxia induces VEGF release

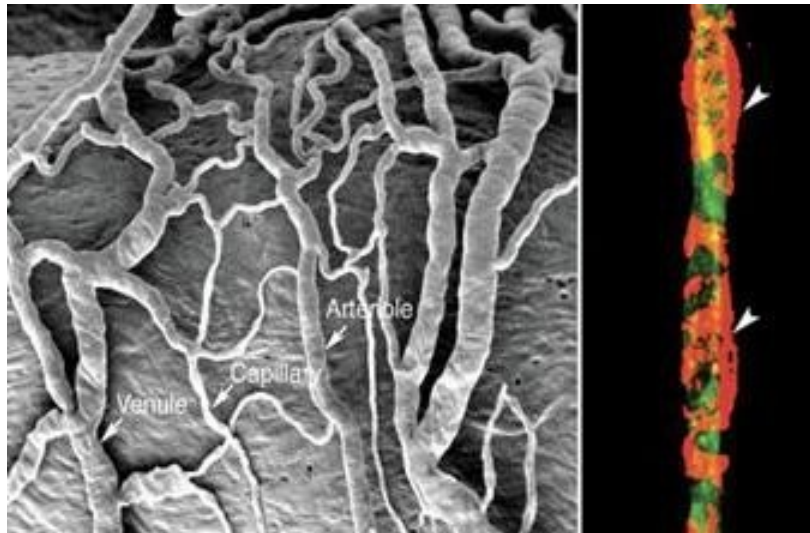
## Vascular growth

### Invasion and metastases



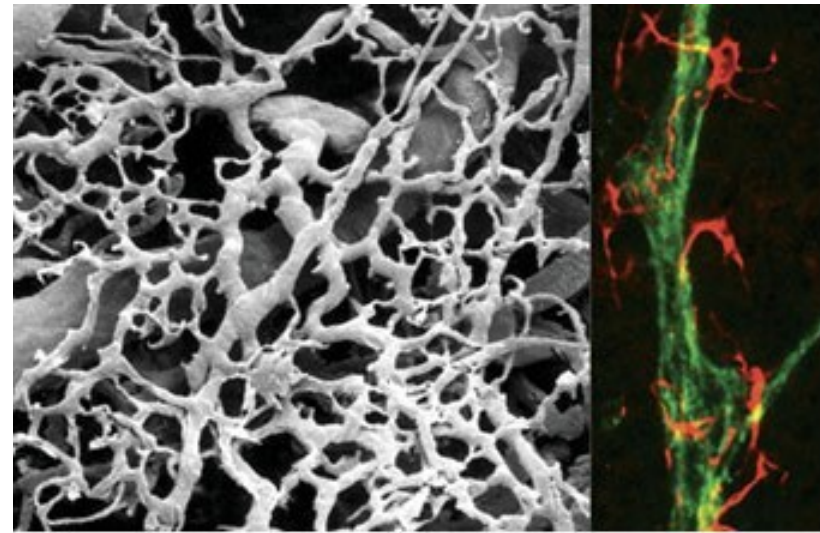
Cells escape from the tumour mass

# Normal vs tumoral vascular network



## Normal network

- organized network
- impermeable vessels
- pericytes coverage (red)



## Tumour network

- disorganized (abnormal, dense and tortuous)
- permeable vessels
- no pericytes coverage (red)

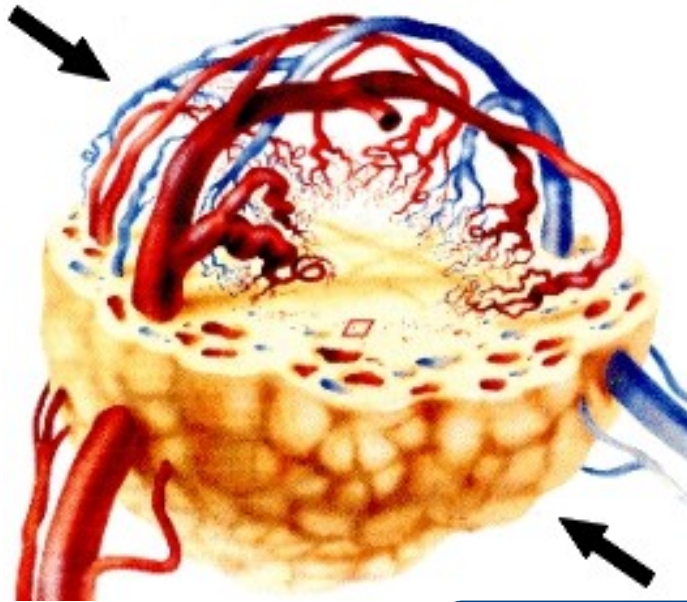


# Therapeutic modes under consideration

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**Antivascular agents**

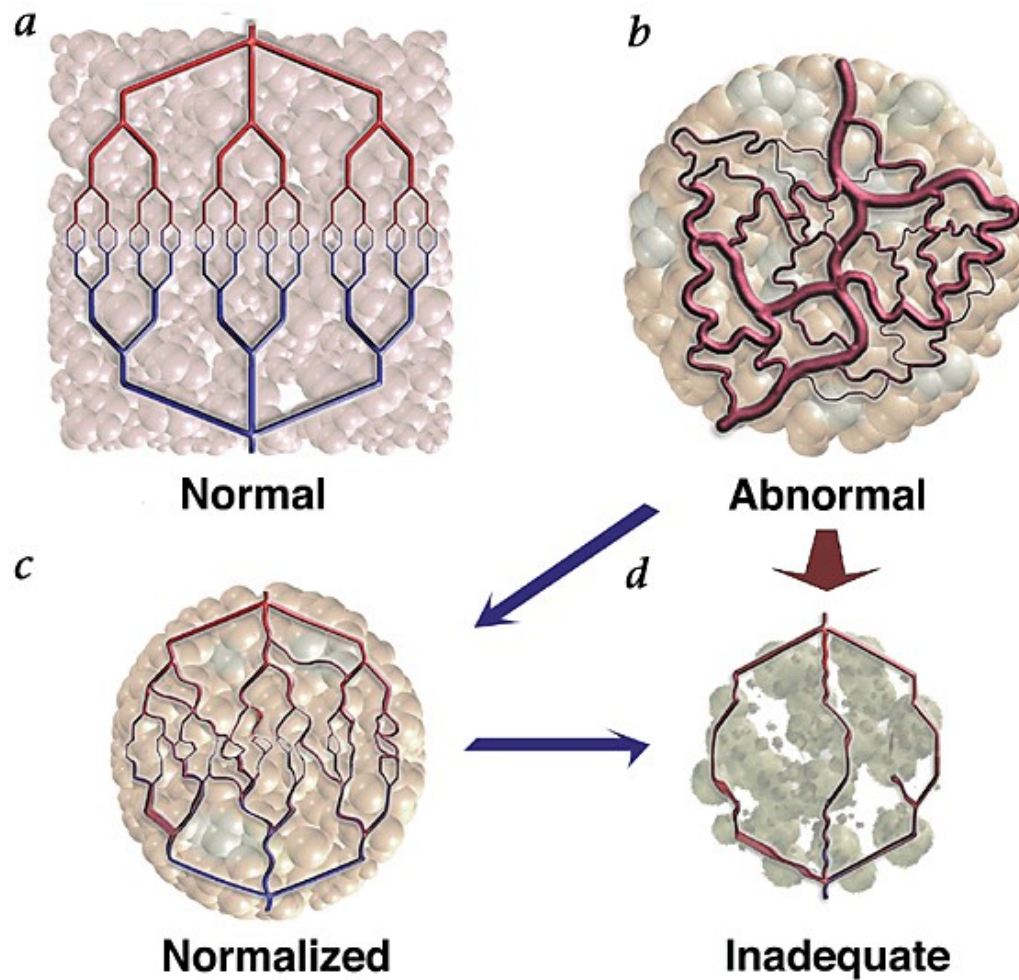
target = vessels



**Cytotoxic molecules**

target = tumour cells

# Therapeutic coupling



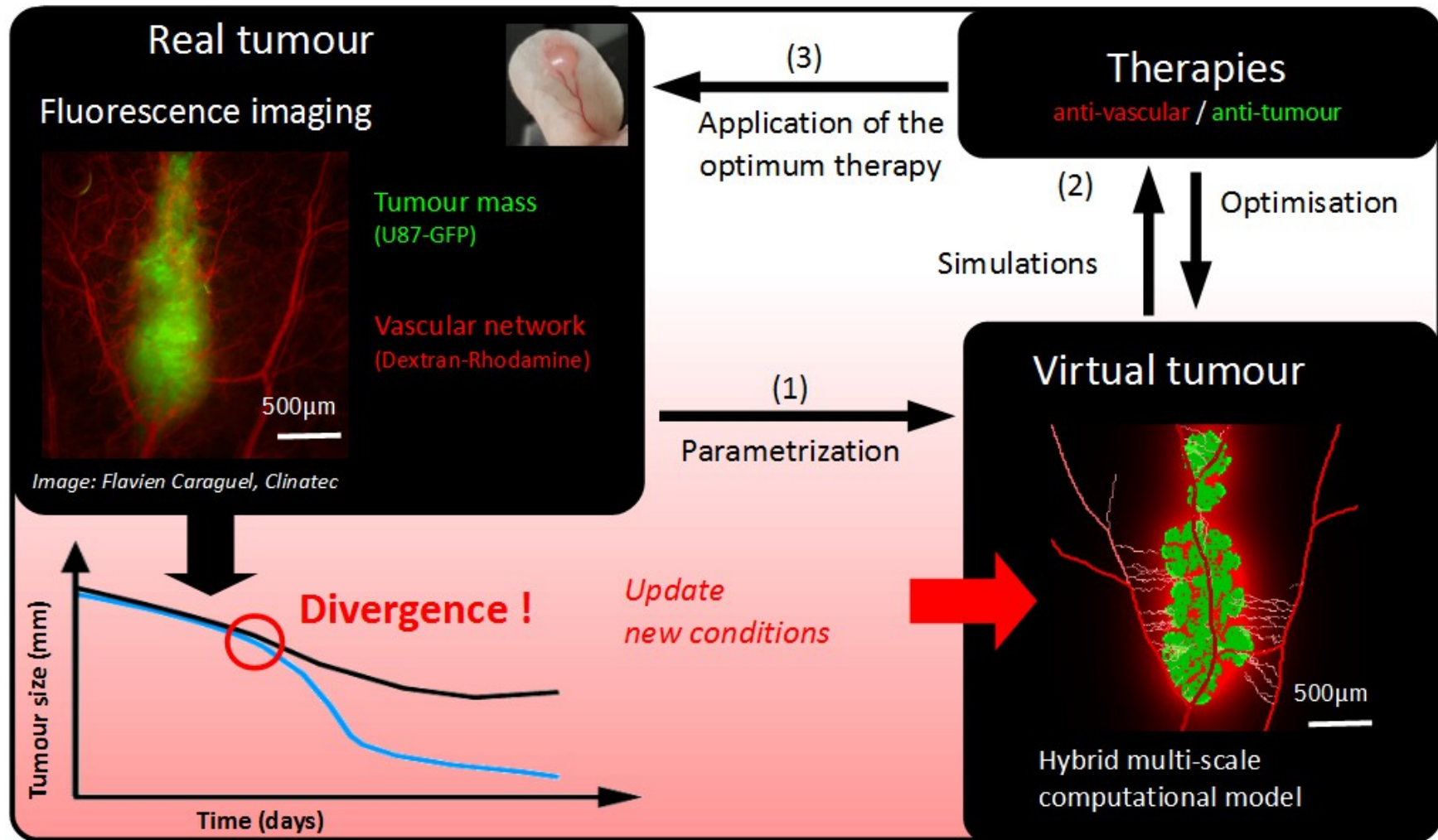
Antivascular agents can **temporarily normalize** the vascular network

The delivery of cytotoxic molecules to the tumour is **transiently improved**

**Therapeutic window**



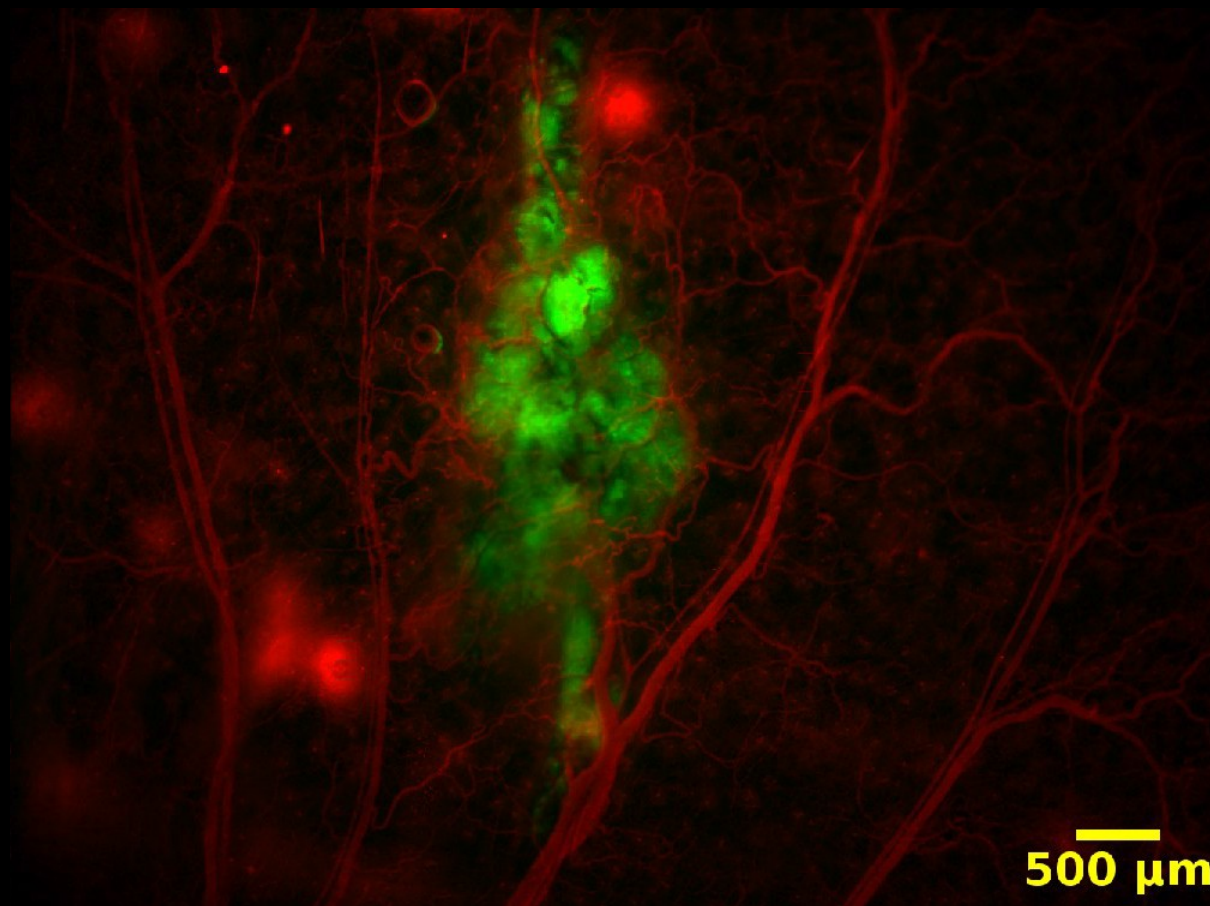
# Computer-assisted therapeutic strategy



# Experimental model



*in vivo* observation through the mouse pinna



**Macrofluorescence  
imaging**

**Tumour mass**  
*(U87-GFP)*

**Vascular network**  
*(Dextran-Rhodamine)*

*Images Flavien Caraguel, Clinattec*

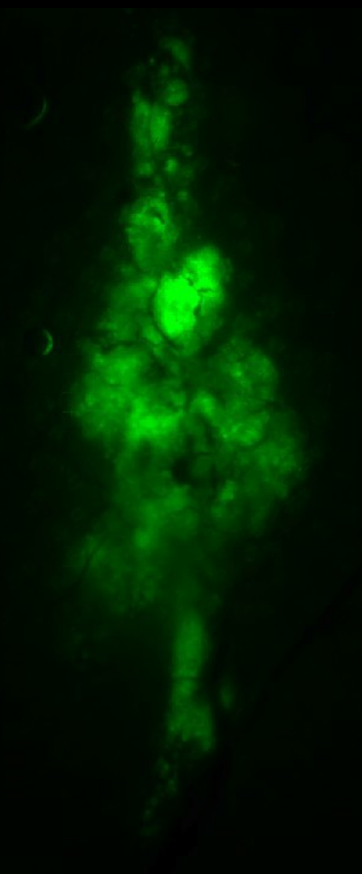
# Experimental model



*in vivo* observation through the mouse pinna

**The two therapeutic modes :**

**Cytotoxic molecules**  
*(target the tumour cells)*



500  $\mu\text{m}$

# Experimental model



*in vivo* observation through the mouse pinna

**The two therapeutic modes :**

Cytotoxic molecules  
*(target the tumour cells)*

Antivascular agents  
*(target the vascular network)*

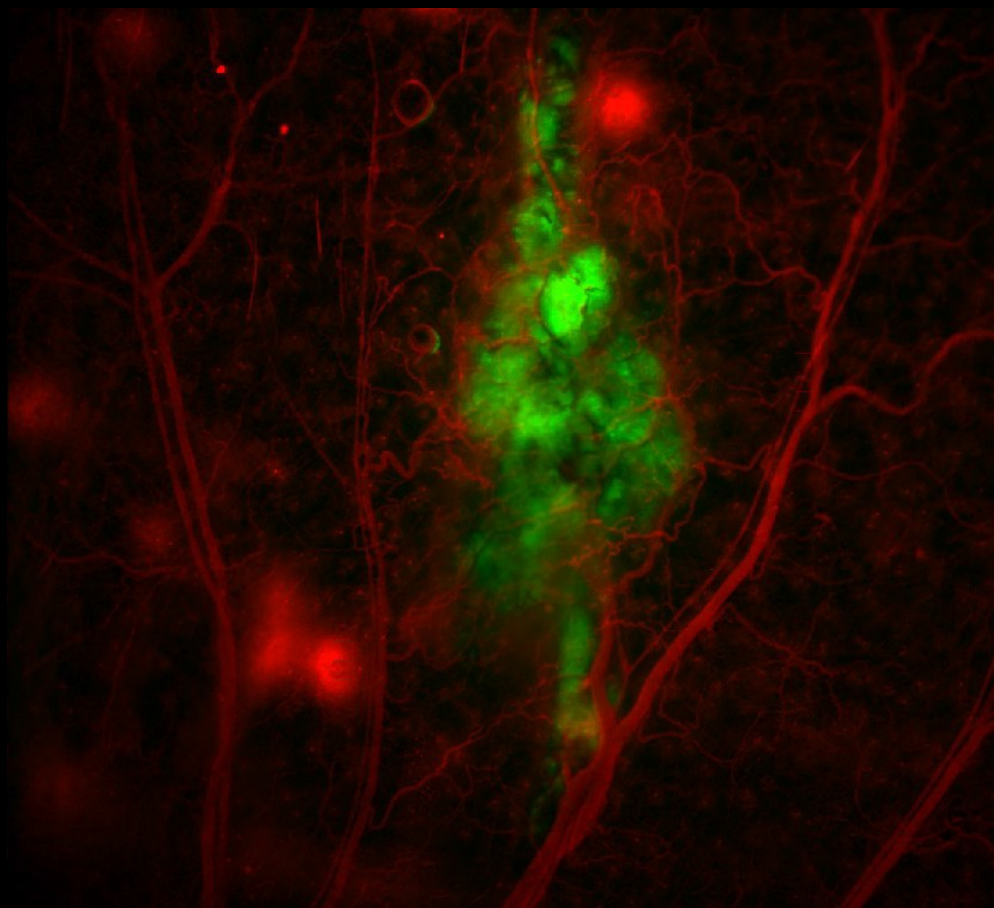
500  $\mu\text{m}$



# Experimental model



*in vivo* observation through the mouse pinna



**The two therapeutic modes :**

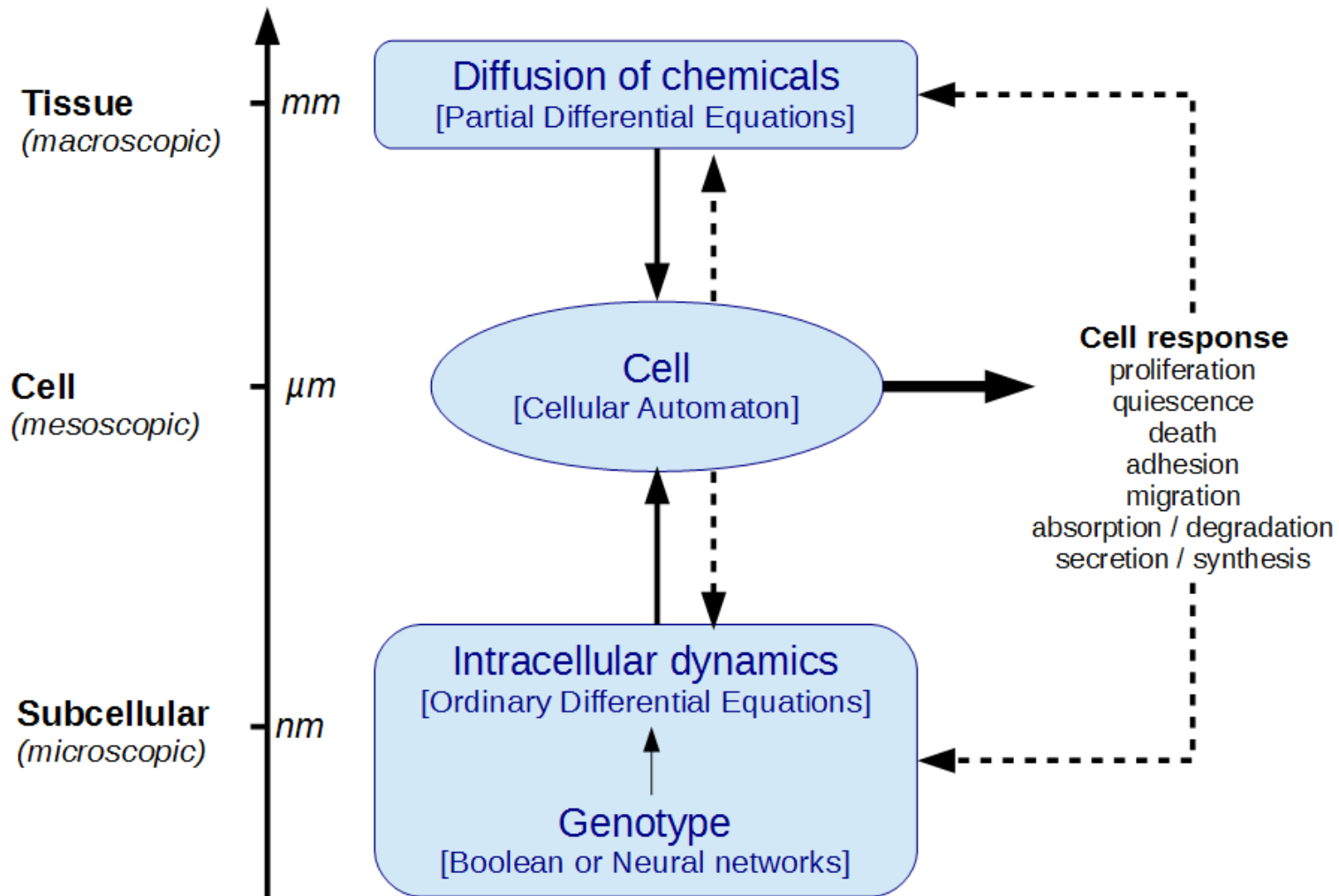
**Cytotoxic molecules**  
*(target the tumour cells)*

+

**Antivascular agents**  
*(target the vascular network)*

**Research of synergetic effects**

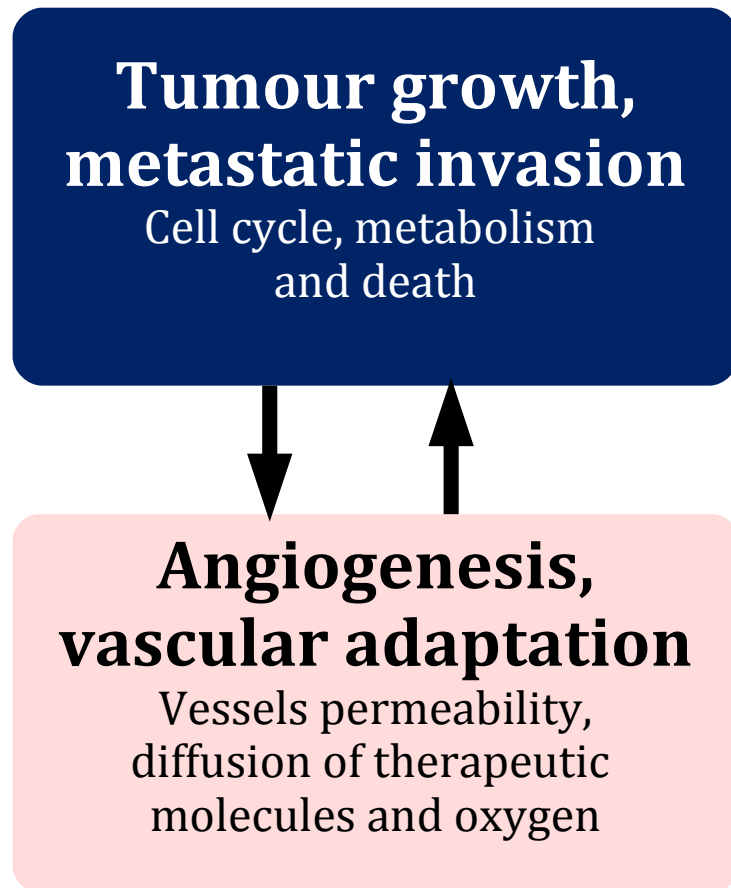
# A cell-centred hybrid multiscale model



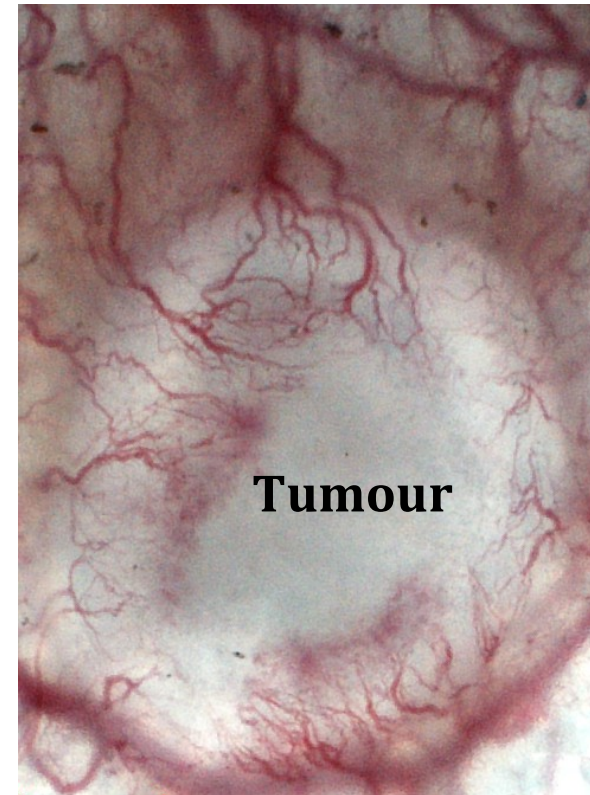


# The computational model

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*Image Ecrins Therapeutics*



1 mm

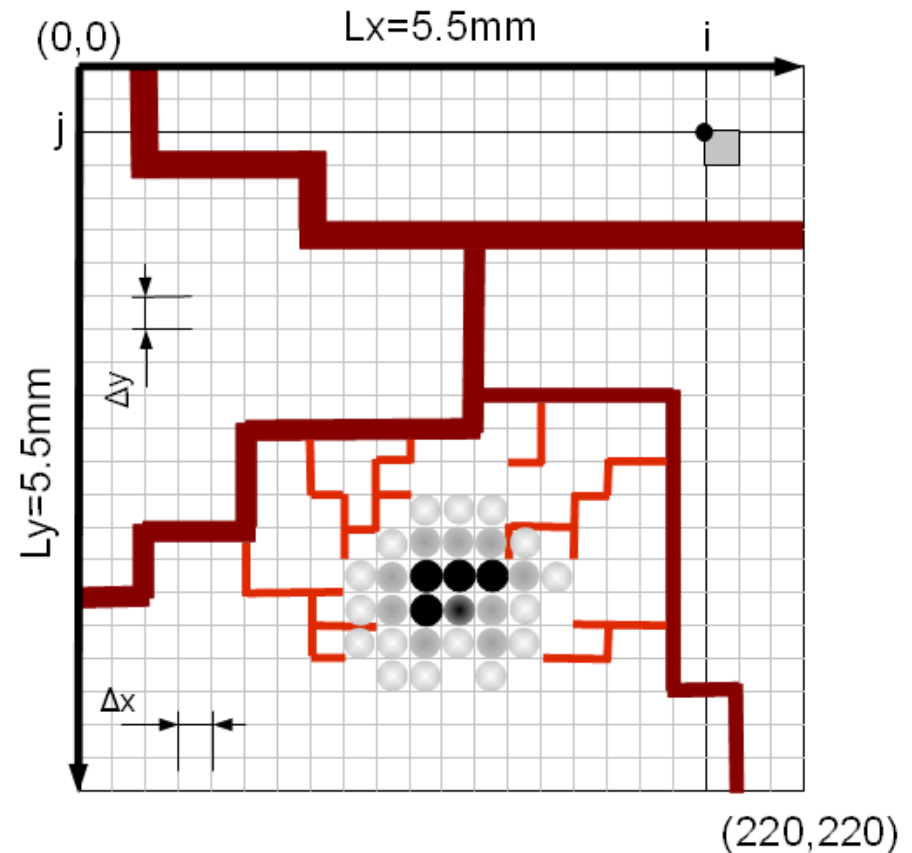
# The computational model

## **Tumour growth, metastatic invasion**

Cell cycle, metabolism  
and death

## **Angiogenesis, vascular adaptation**

Vessels permeability,  
diffusion of therapeutic  
molecules and oxygen



# Diffusive species

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## Proteases

Matrix degrading enzyme ( $m$ )

$$\frac{\partial m}{\partial t} = D_m \nabla^2 m + \alpha_m \textcolor{red}{n}_{i,j} - \nu_m m$$

*Endothelial cell*

Vascular degrading enzyme ( $p$ )

$$\frac{\partial p}{\partial t} = D_p \nabla^2 p + \alpha_p \textcolor{red}{P}_{i,j} - \nu_p p$$

*Proliferative cell*

## Growth factors ( $V$ )

$$\frac{\partial V}{\partial t} = D_V \nabla^2 V + \alpha_V \textcolor{red}{Q}_{i,j} - \nu_V V - \lambda_V \textcolor{blue}{W}_{i,j} \min(V, V_{max})$$

*Quiescent cell*

*Vessels "weight"*

## Oxygen ( $O$ )

$$\frac{\partial O}{\partial t} = D_O \nabla^2 O + \gamma_v \textcolor{blue}{W}_{i,j} (O_v - O) - \textcolor{red}{k}_{i,j} O$$

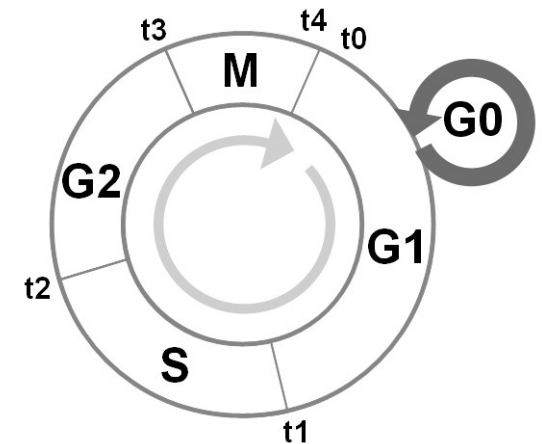
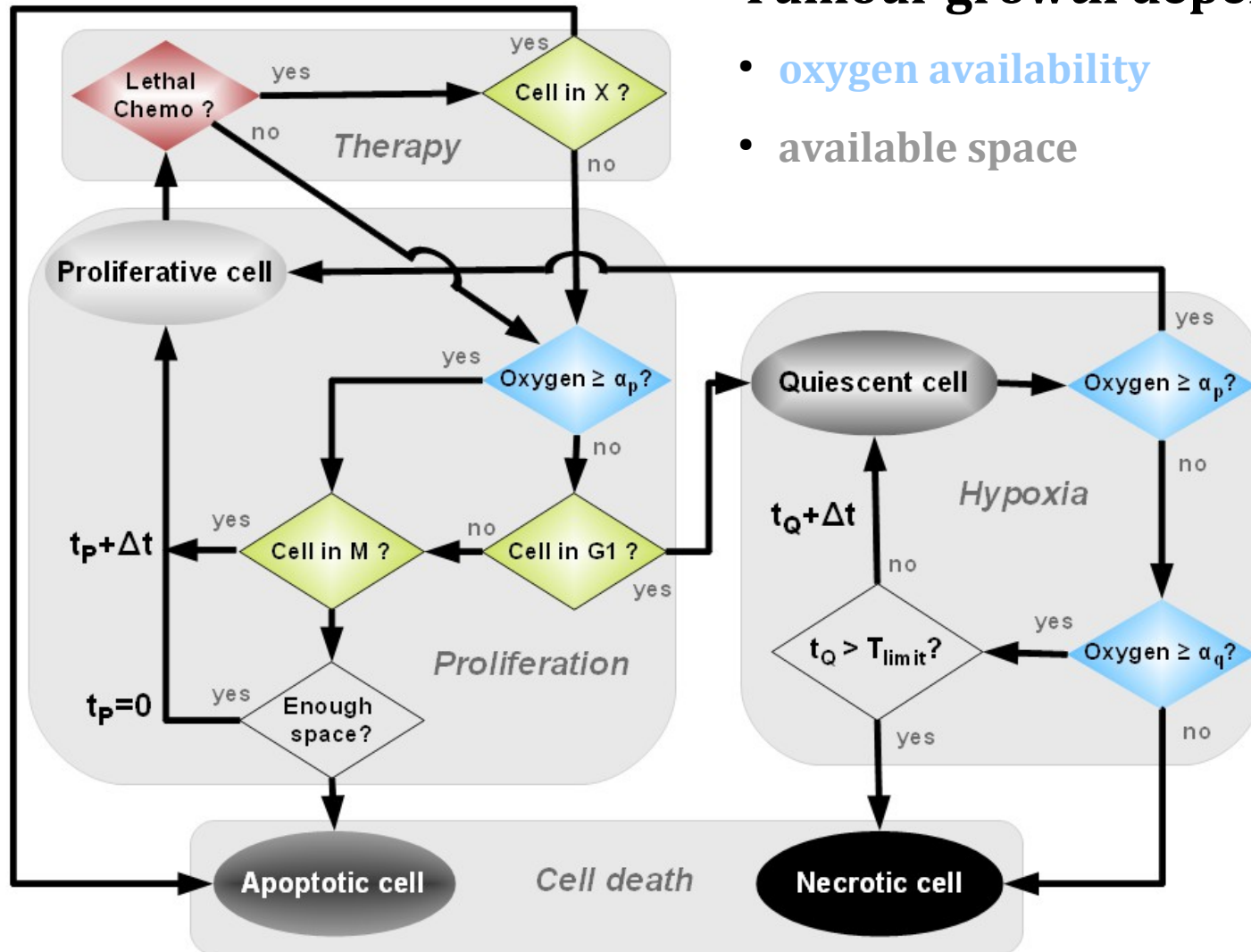
*Vessels "weight"*

*kn, kp, kq*

# Modelling tumour growth

Tumour growth depends on:

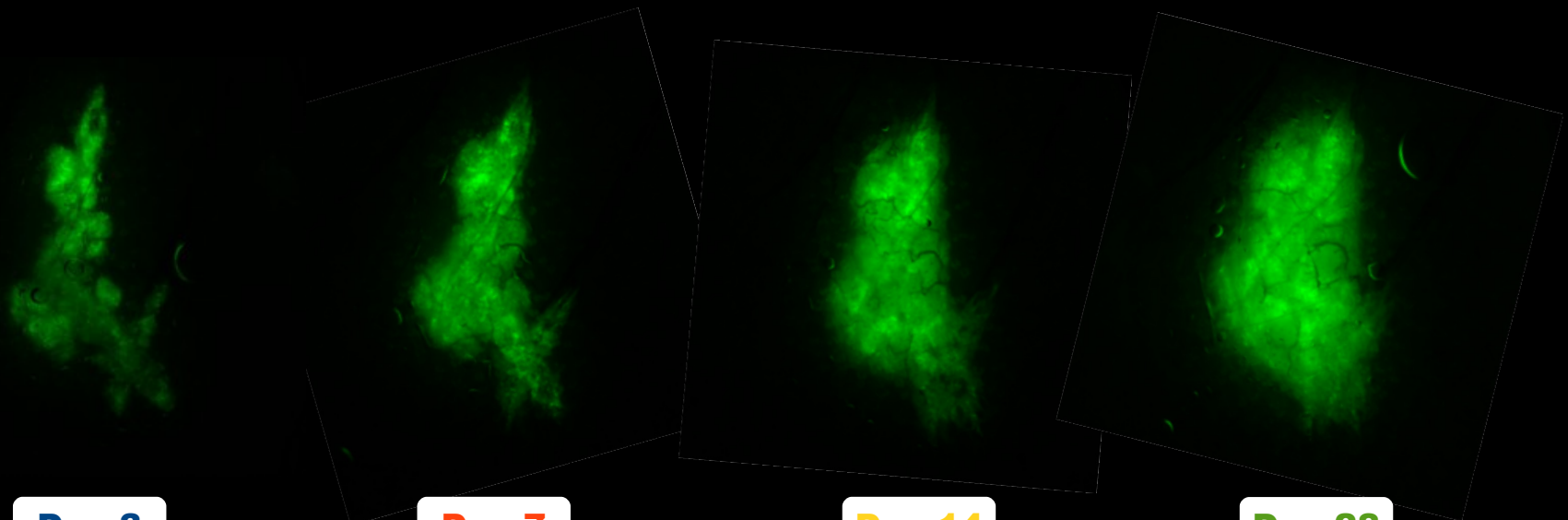
- oxygen availability
- available space
- cell cycle
- concentration of cytotoxic molecules



# Tumour growth : texture and size

**Real  
tumour**

U87-GFP

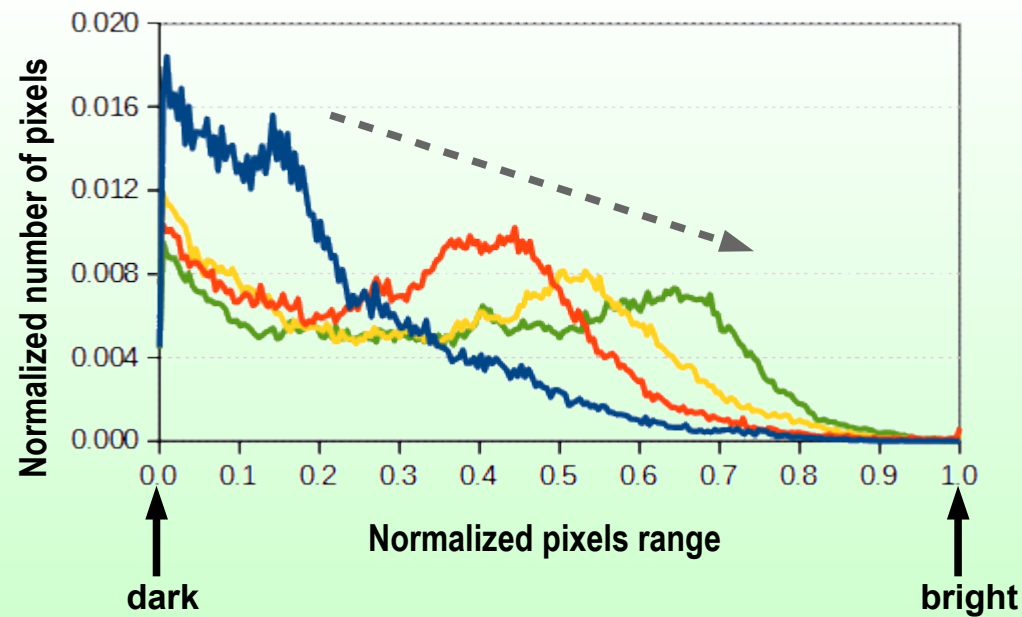


**Day 3**

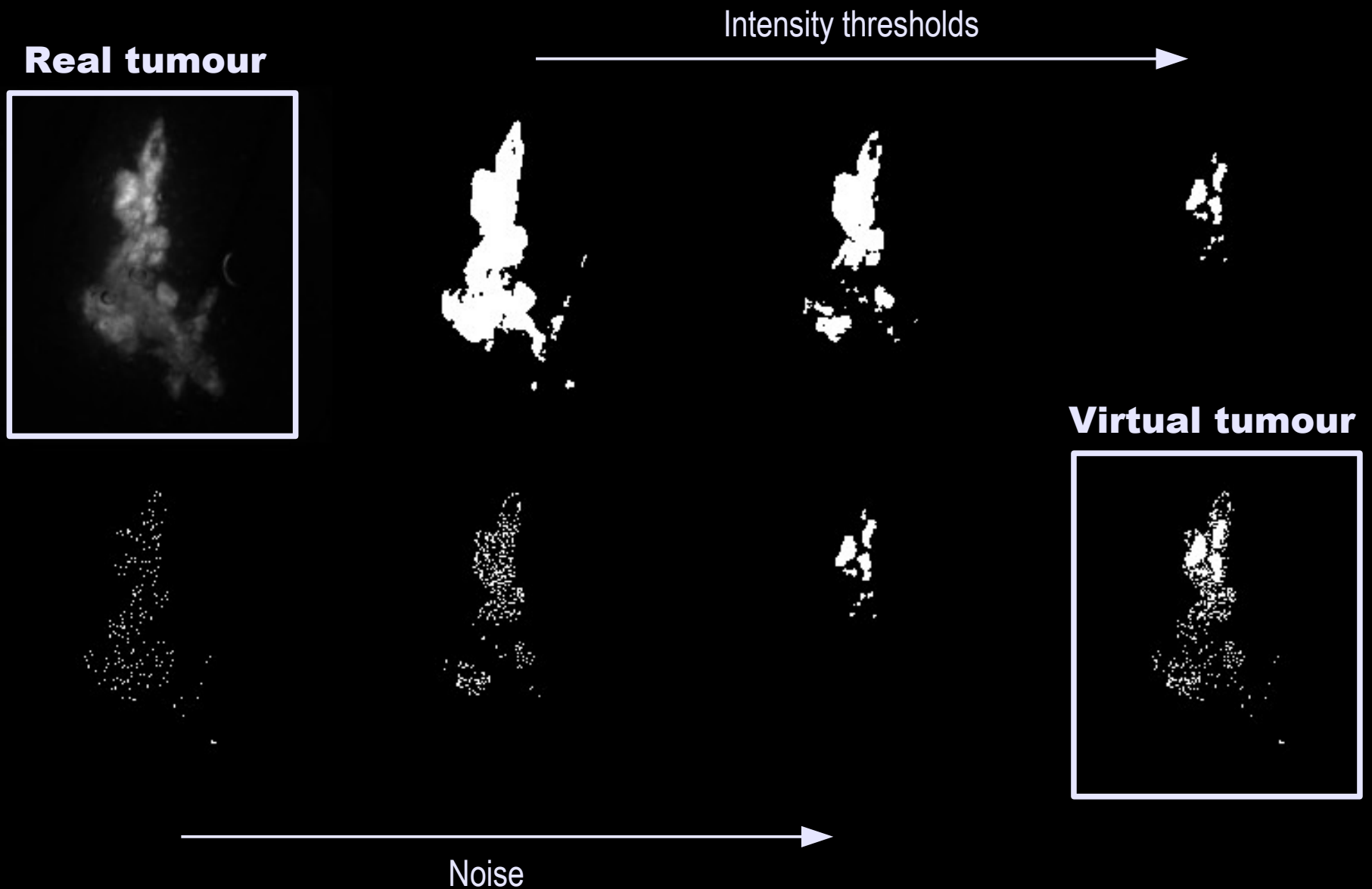
**Day 7**

**Day 14**

**Day 28**



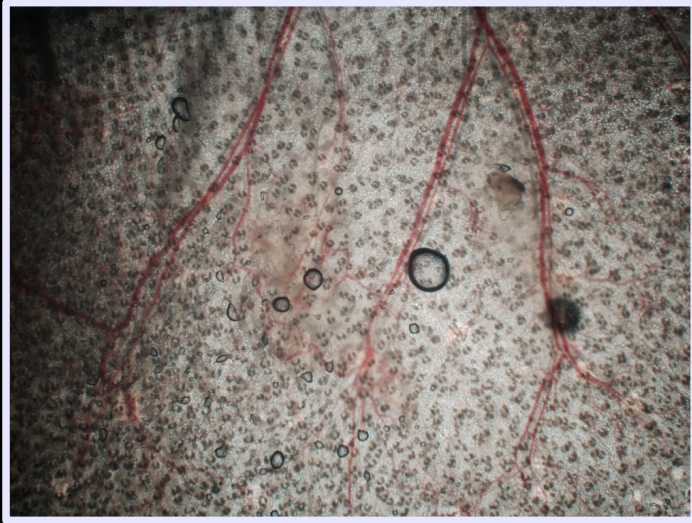
# Initialization of the tumour



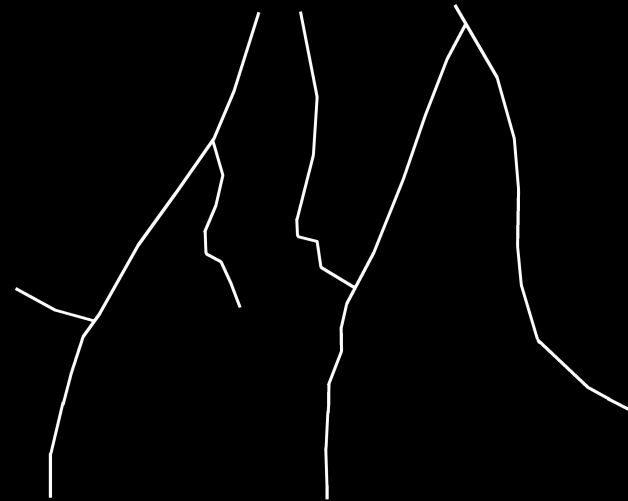


# Initialization of the tumour

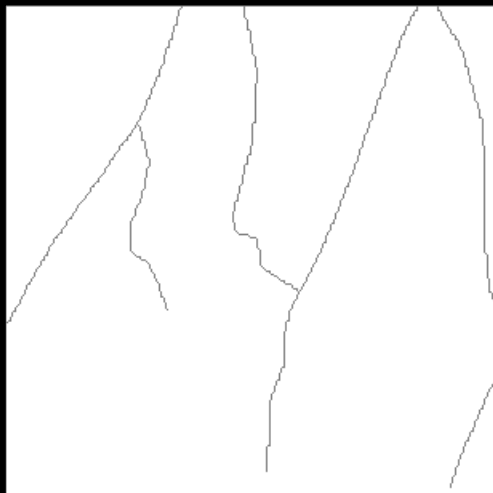
**Bright field image**



Vascular skeleton



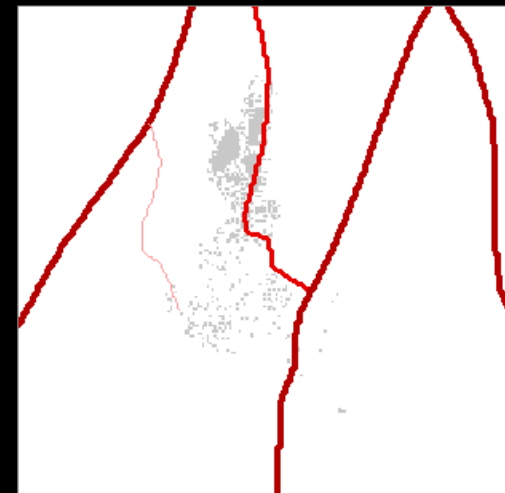
Cropped image



Perfusion



Initial vasculature + tumour



# Real vs virtual tumour

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Day 3

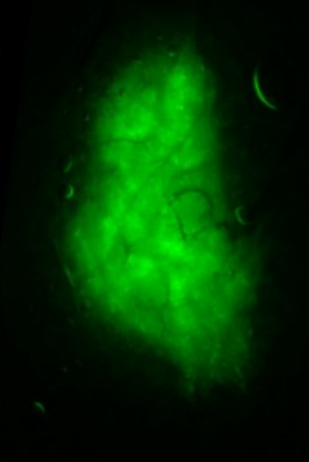
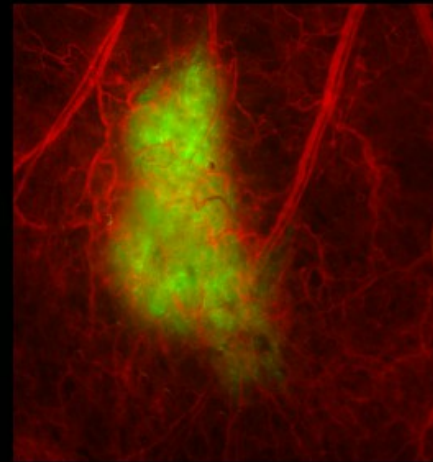
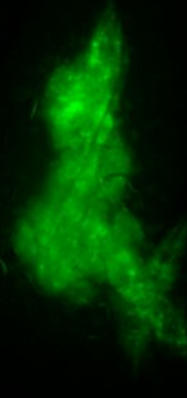
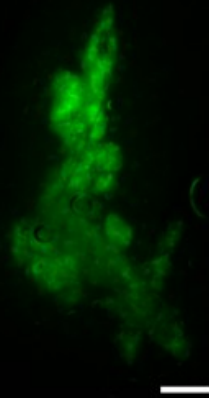
Day 7

Day 14

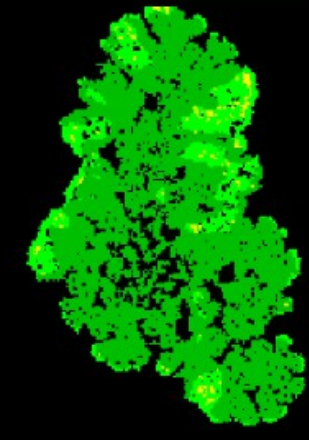
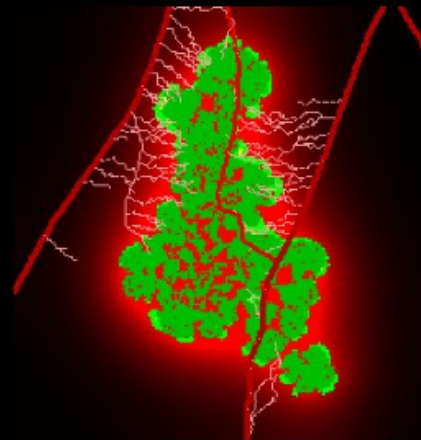
Day 28

**Real  
tumour**

U87-GFP



**Virtual  
tumour**



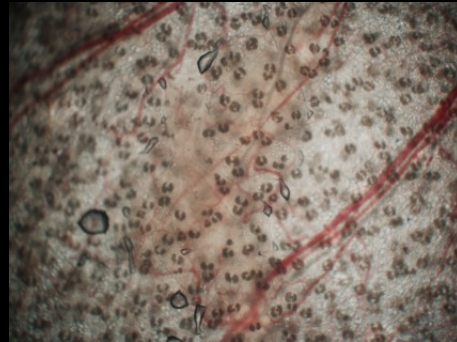
# Real vs virtual tumour

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**Day 3**



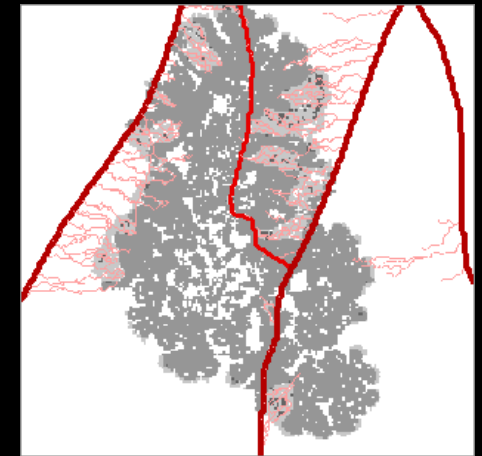
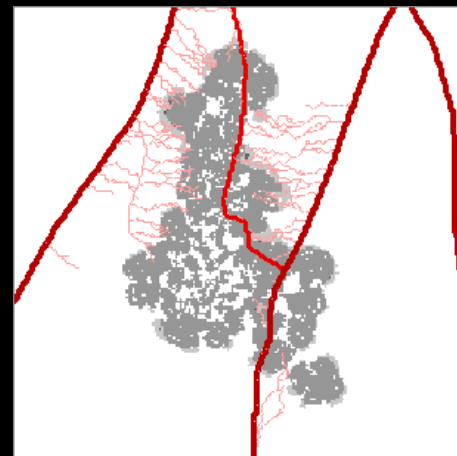
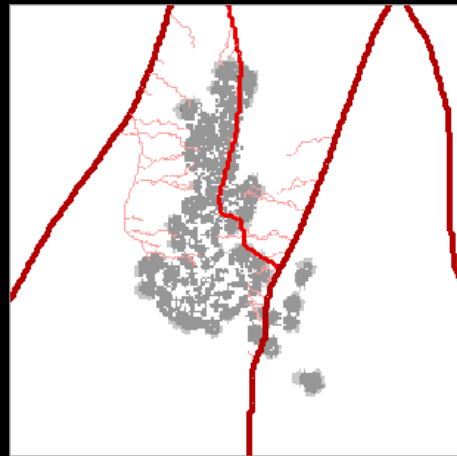
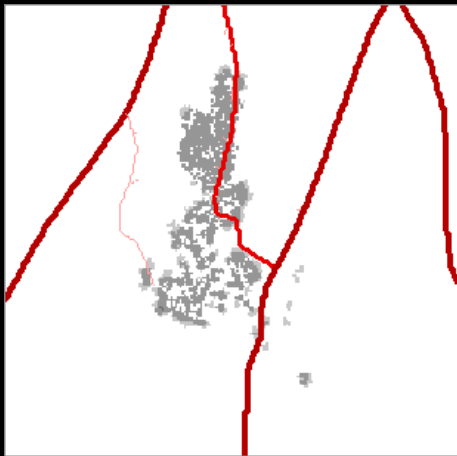
**Day 7**



**Day 14**



**Day 28**



**Angiogenesis**

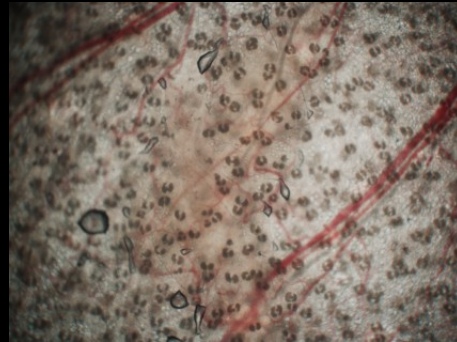


# Real vs virtual tumour

Day 3



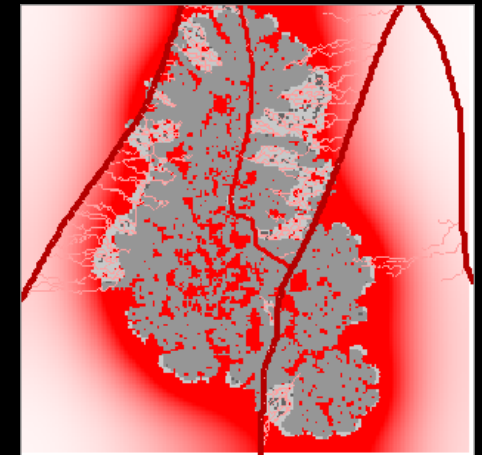
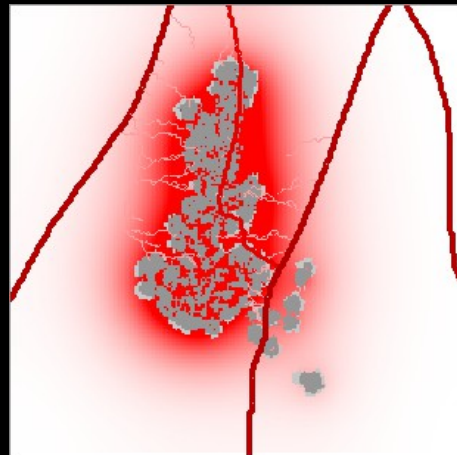
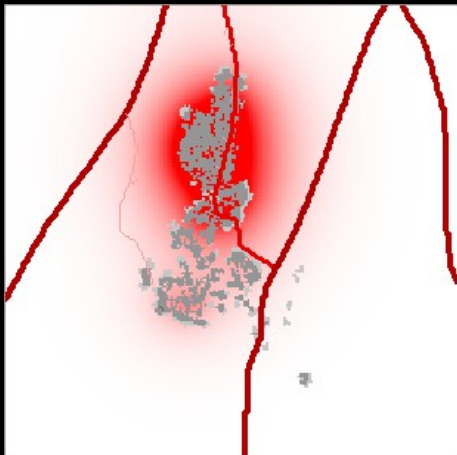
Day 7



Day 14



Day 28

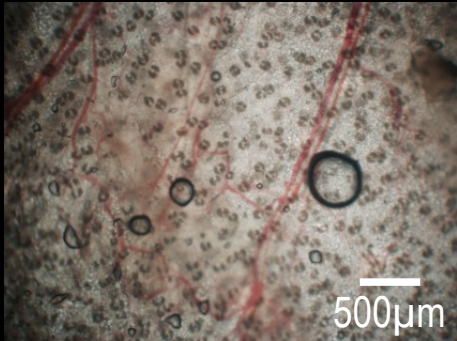


**Angiogenesis + VEGF diffusion**

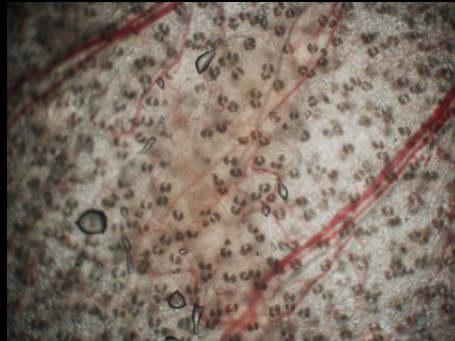
# Real vs virtual tumour

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**Day 3**



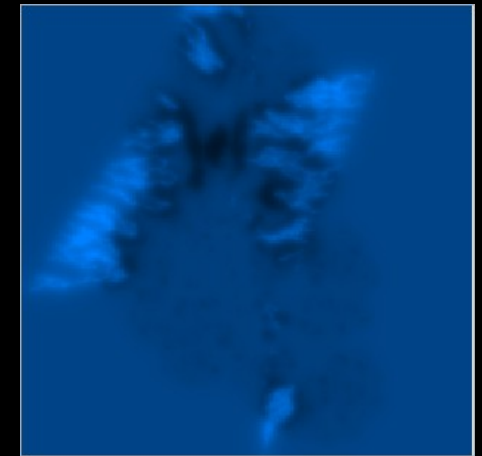
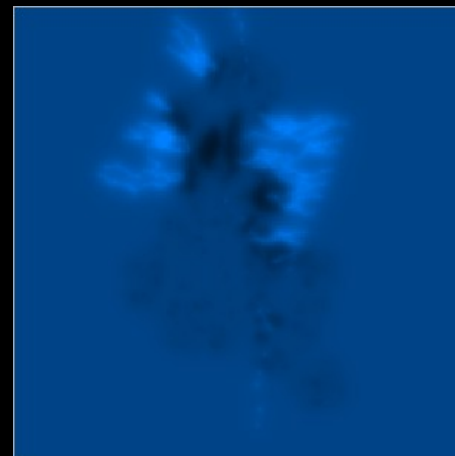
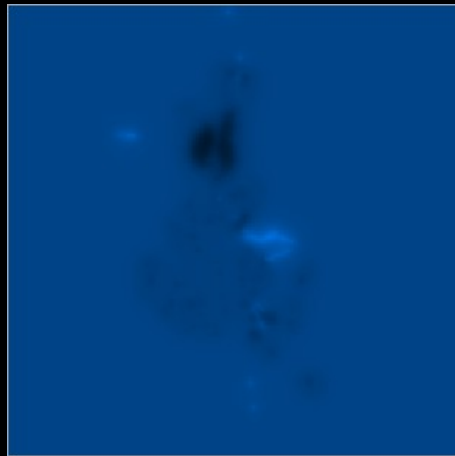
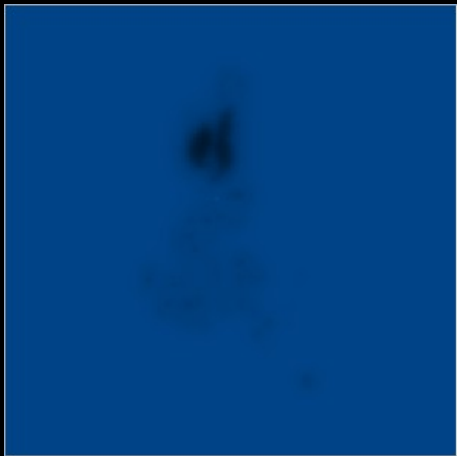
**Day 7**



**Day 14**

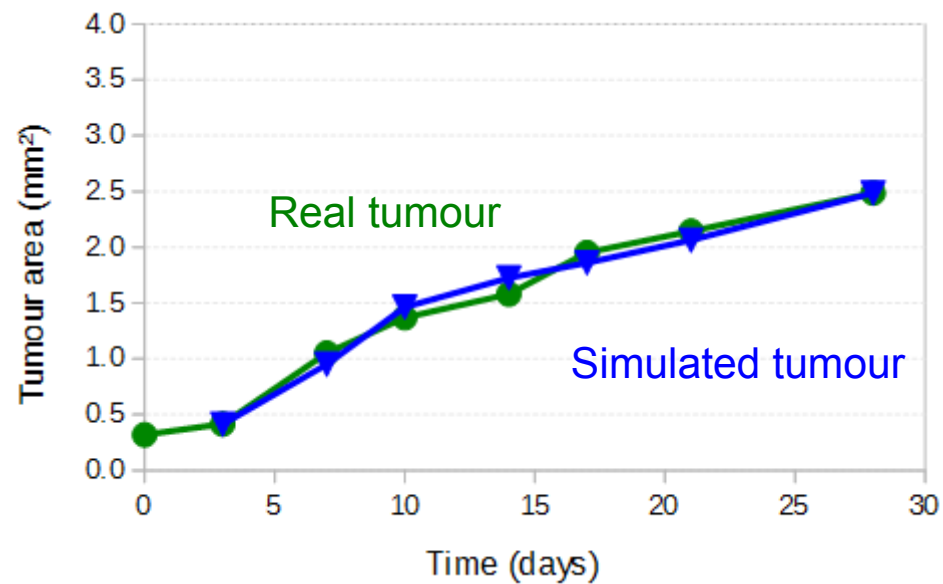
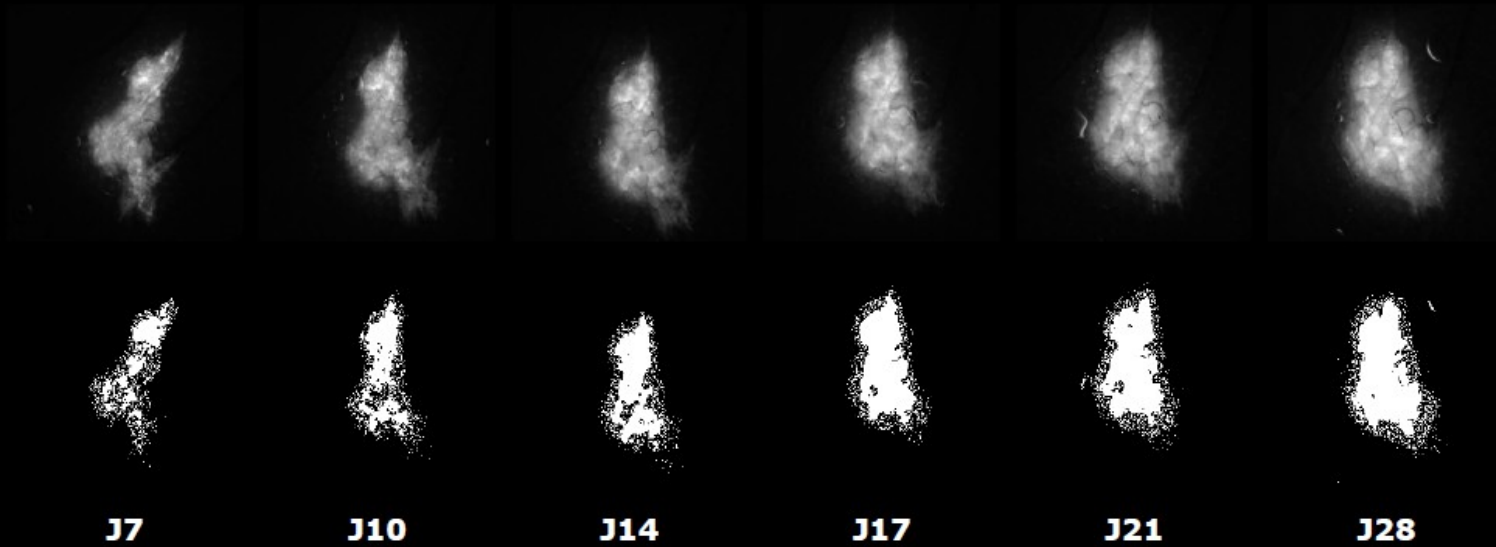


**Day 28**



**Oxygen diffusion**

# Growth without treatment





# Model validation

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- ✓ Comparison of simulated and observed kinetics of tumour development
- ✓ Comparison of virtual and histological tumour slices
- ✓ **Adjustment of parameters and validation of hypotheses**

## **Parameters adjusted**

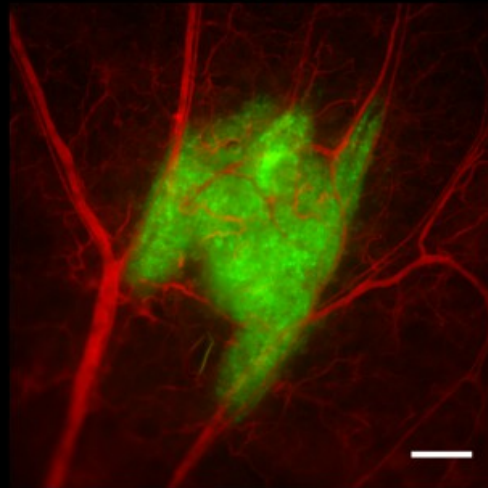
- Durations of cell cycle phases and level of variability
- **Oxygen thresholds for transition to quiescence (hypoxia) and for cell death**
- Oxygen consumption rates for each cell type
- Vessels permeability to oxygen
- Diffusion, production/consumption rates of vascular proteases

# Real vs virtual tumour

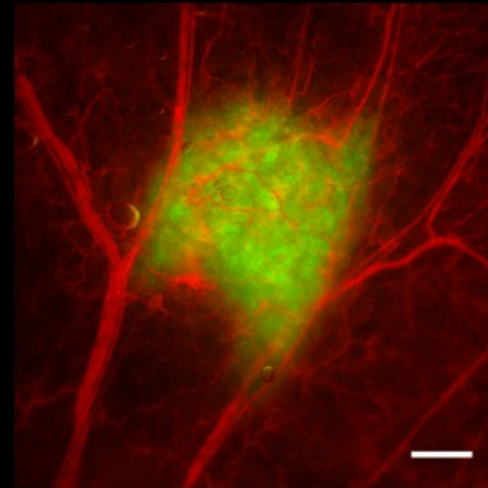
**Real  
Tumour**

**Experiment**

**Day 7**



**Day 14**

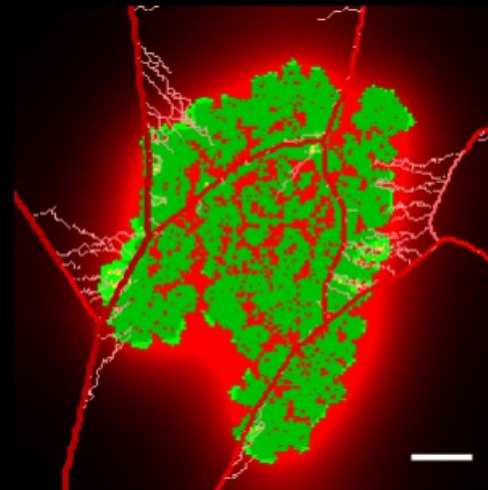
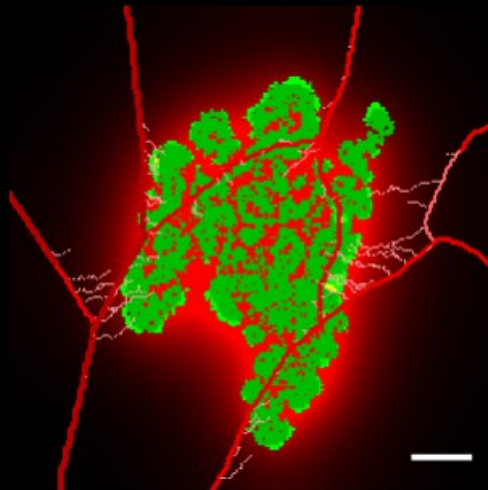


**U87-GFP**

**Dextran-rhodamine**

**Virtual  
Tumour**

**Simulation**

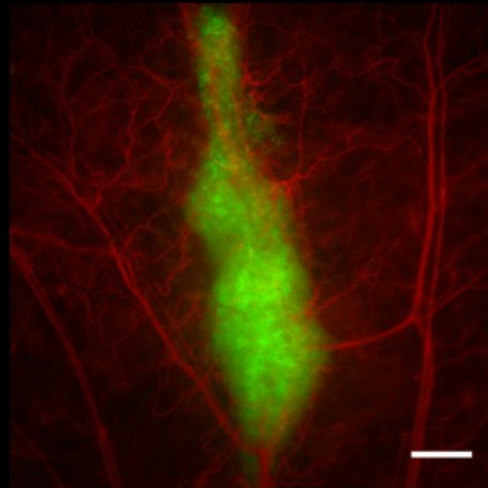


# Real vs virtual tumour

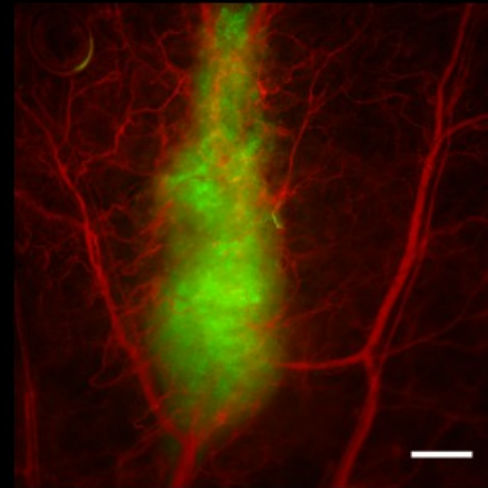
**Real  
Tumour**

**Experiment**

**Day 7**



**Day 14**

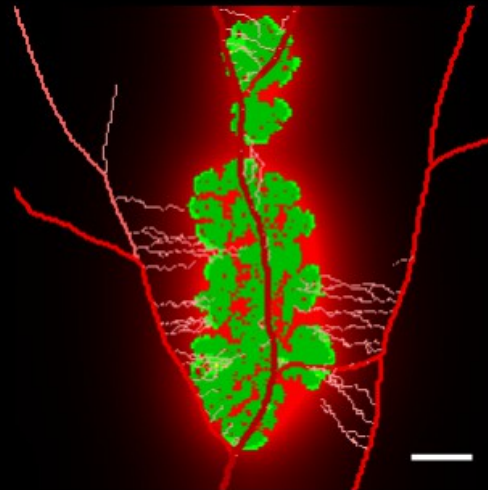
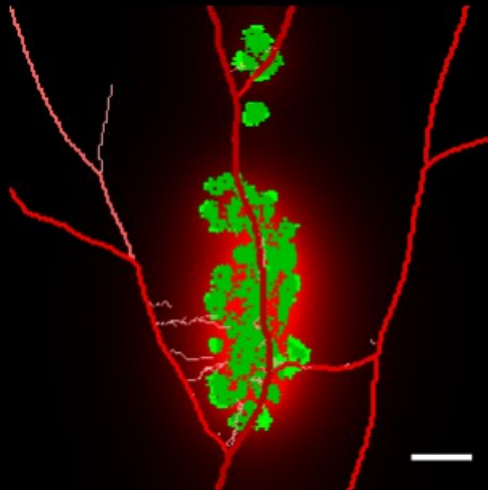


**U87-GFP**

**Dextran-rhodamine**

**Virtual  
Tumour**

**Simulation**

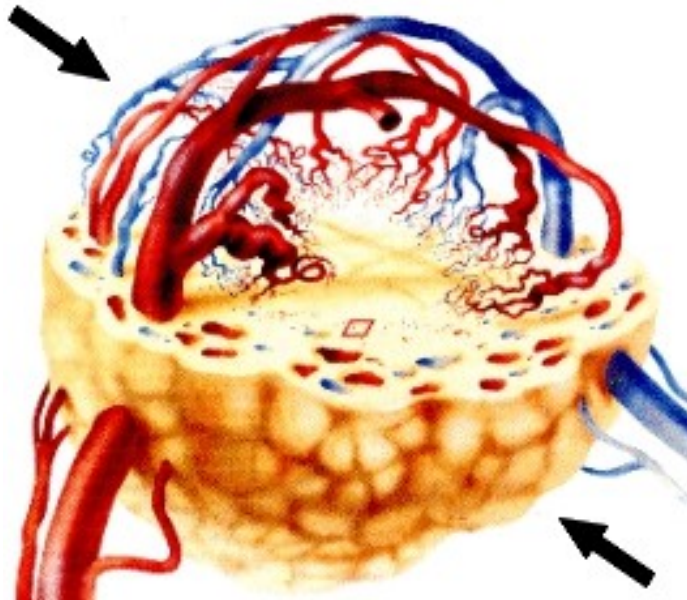


# Therapies

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**Antivascular agents**

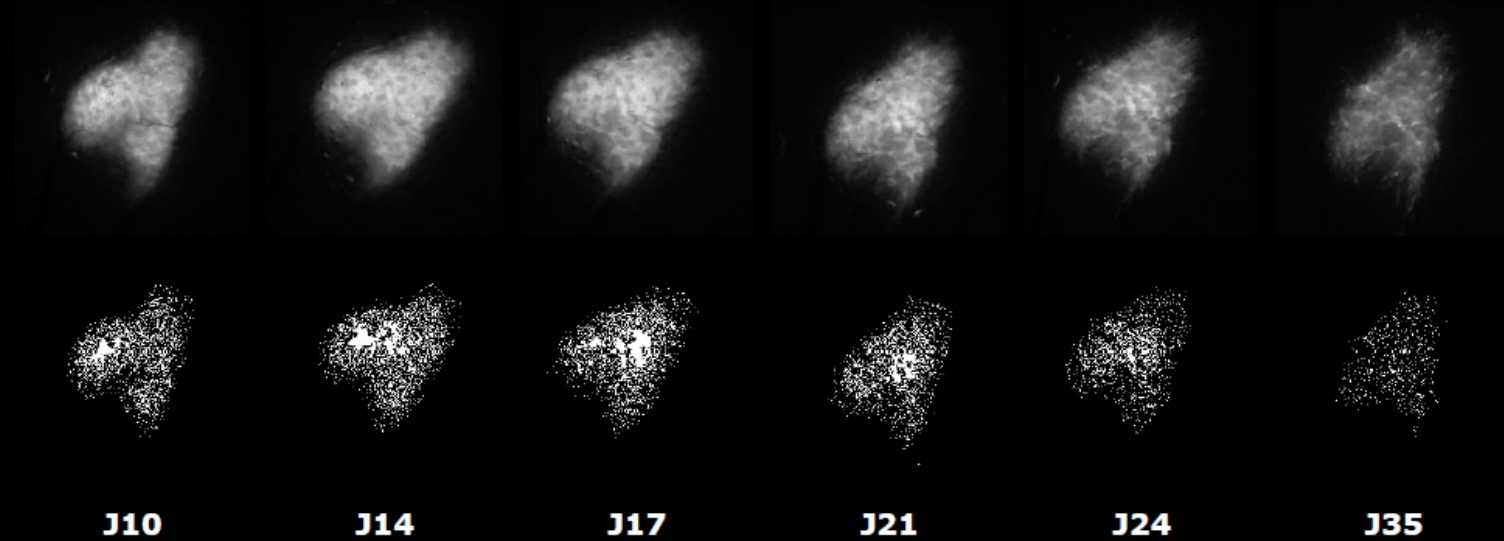
target = vessels



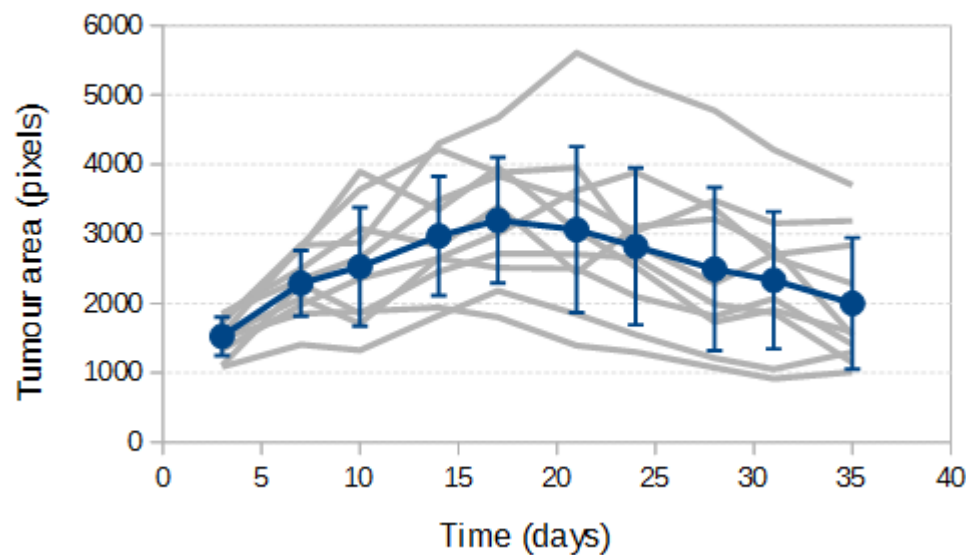
**Cytotoxic molecules**

target = tumour cells

# Growth with cytotoxic treatment (temodal)



Average evolution  
from 12 tumours

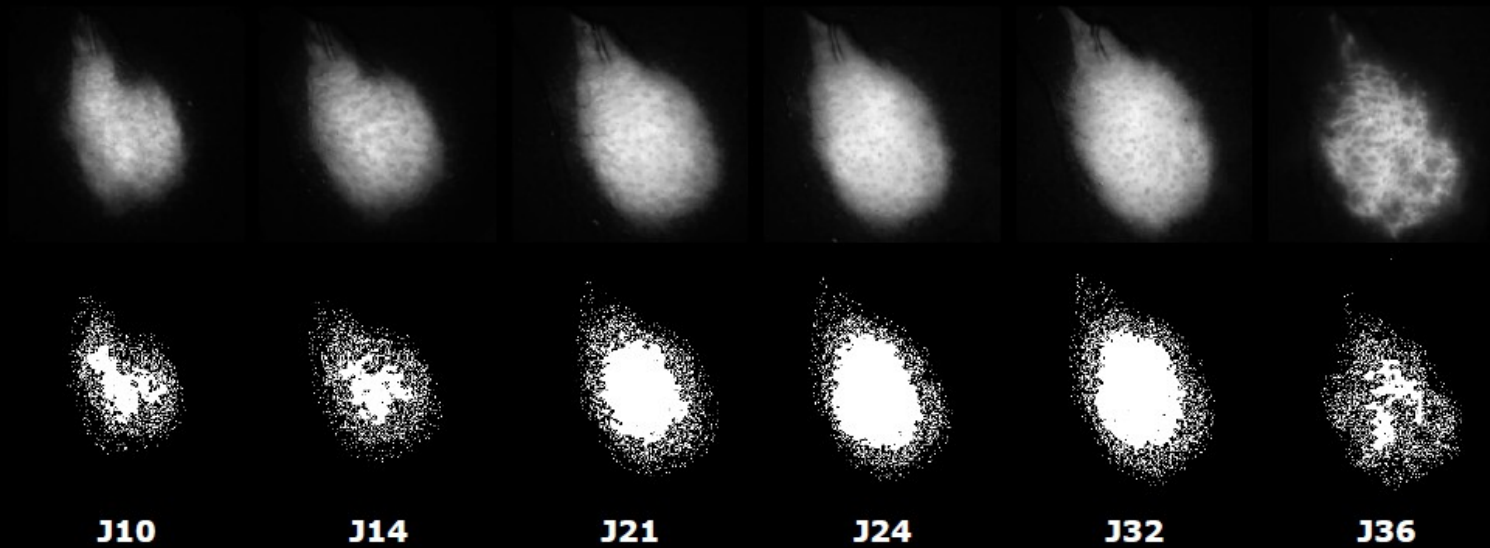


## Dosage

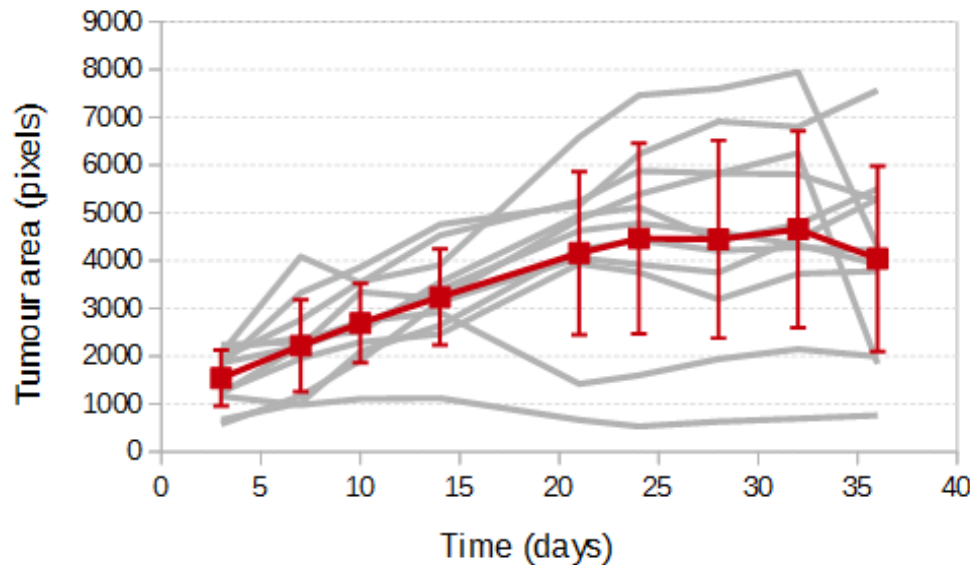
50mg/kg (in food)

5 consecutive days  
(days 14 to 18)

# Growth with antivasclar treatment (avastin)



Average evolution  
from 12 tumours



## Dosage

10mg/kg (IP injection)

Every 3 days in average  
(days 14, 17, 22, 25, 28)

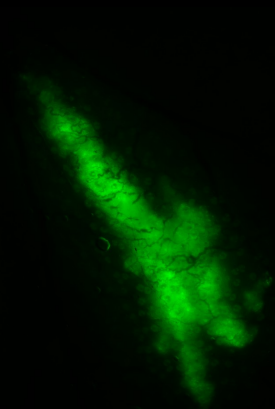


# Tumour case 1 (highly vascularized)

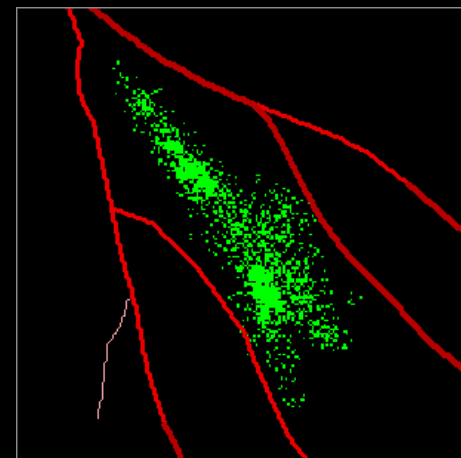
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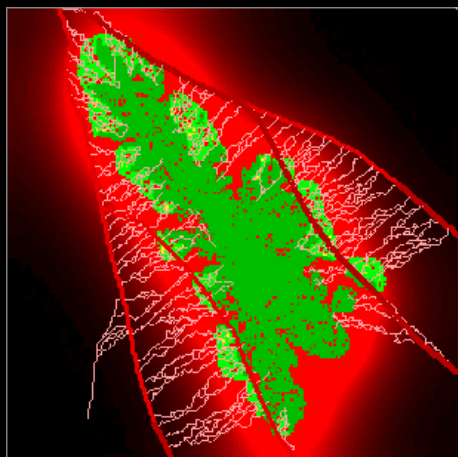
Vascular network (day 3)



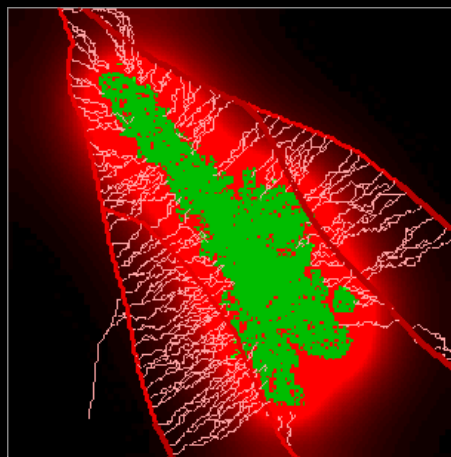
GFP-Image (day3)



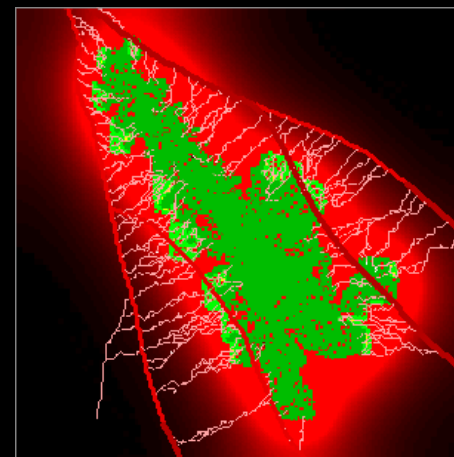
Simulation (day 3)



Simulation without  
treatment (day 34)



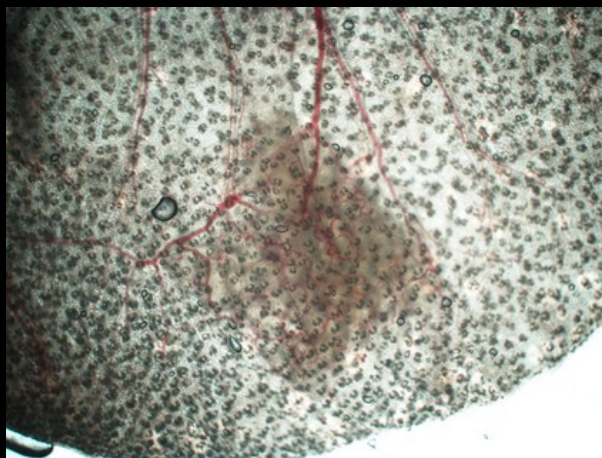
Simulation with  
temodal (day 34)



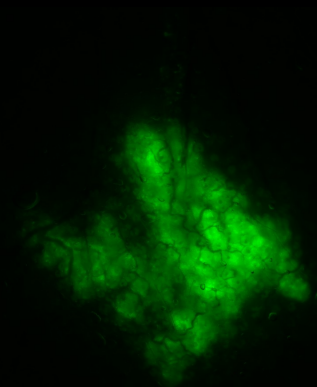
Simulation with  
avastin (day 34)

# Tumour case 2 (weakly vascularized)

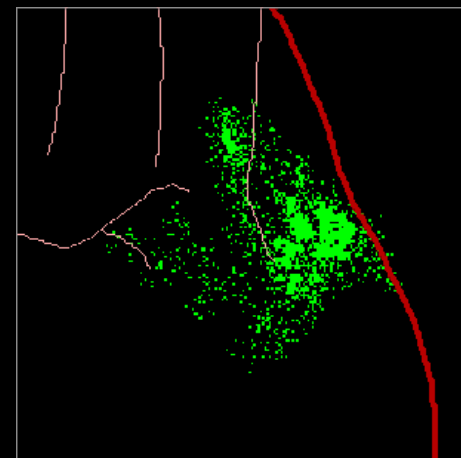
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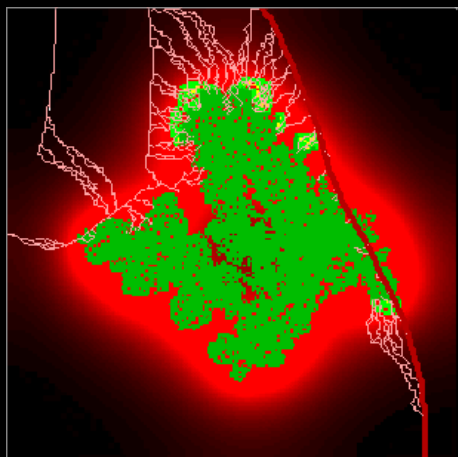
Vascular network (day 3)



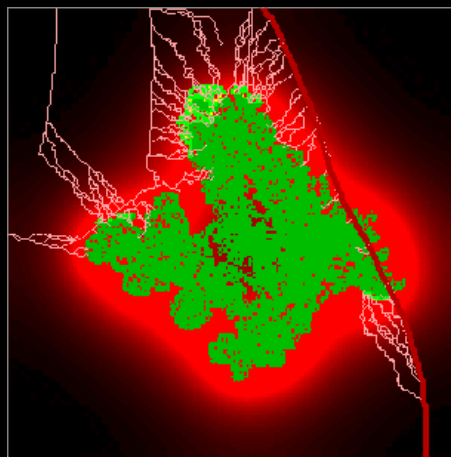
GFP-Image (day3)



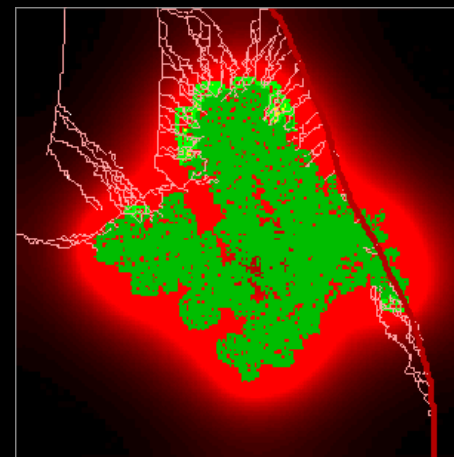
Simulation (day 3)



Simulation without  
treatment (day 34)



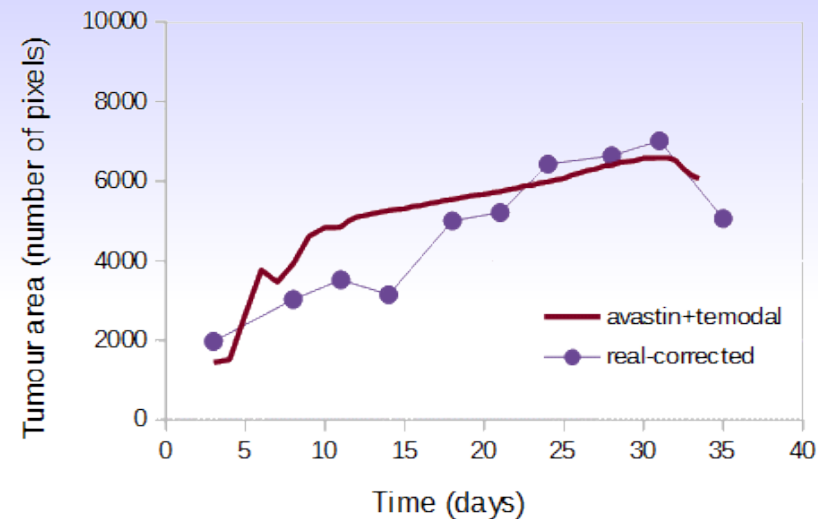
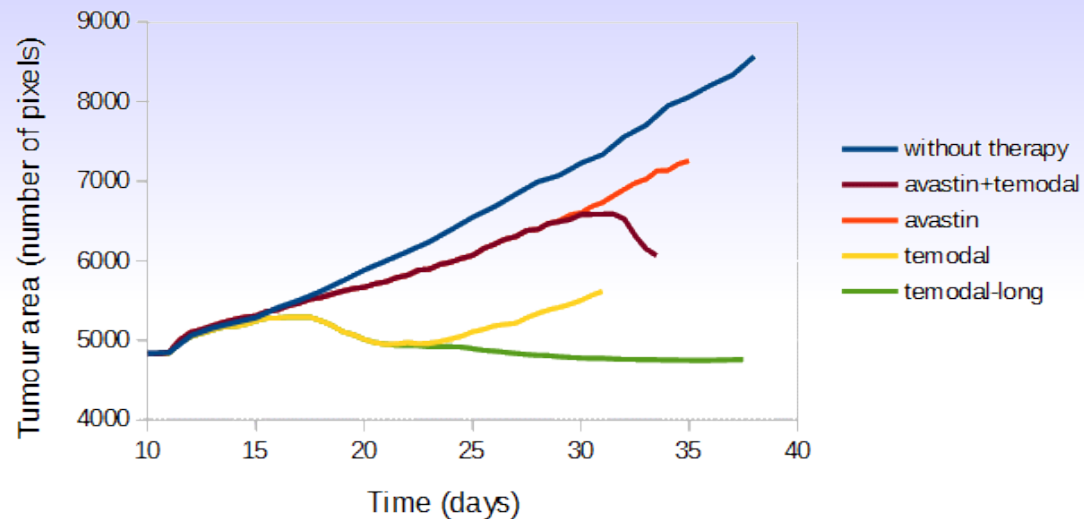
Simulation with  
temodal (day 34)



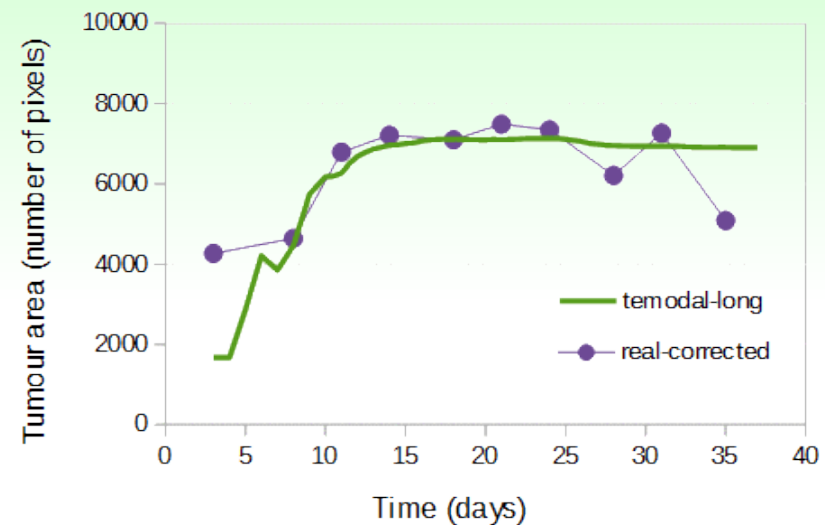
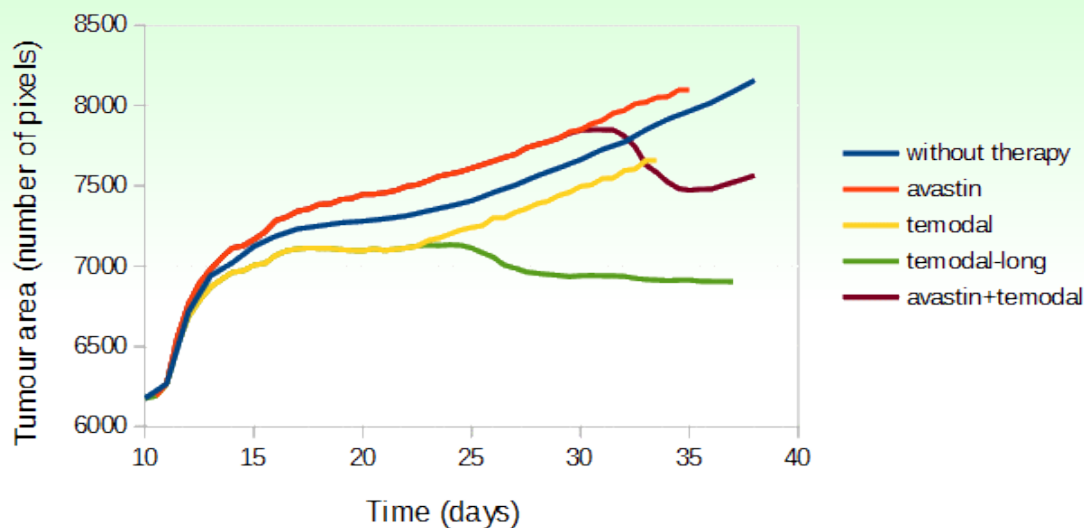
Simulation with  
avastin (day 34)

# Predictions

## Tumour case 1 (highly vascularized)

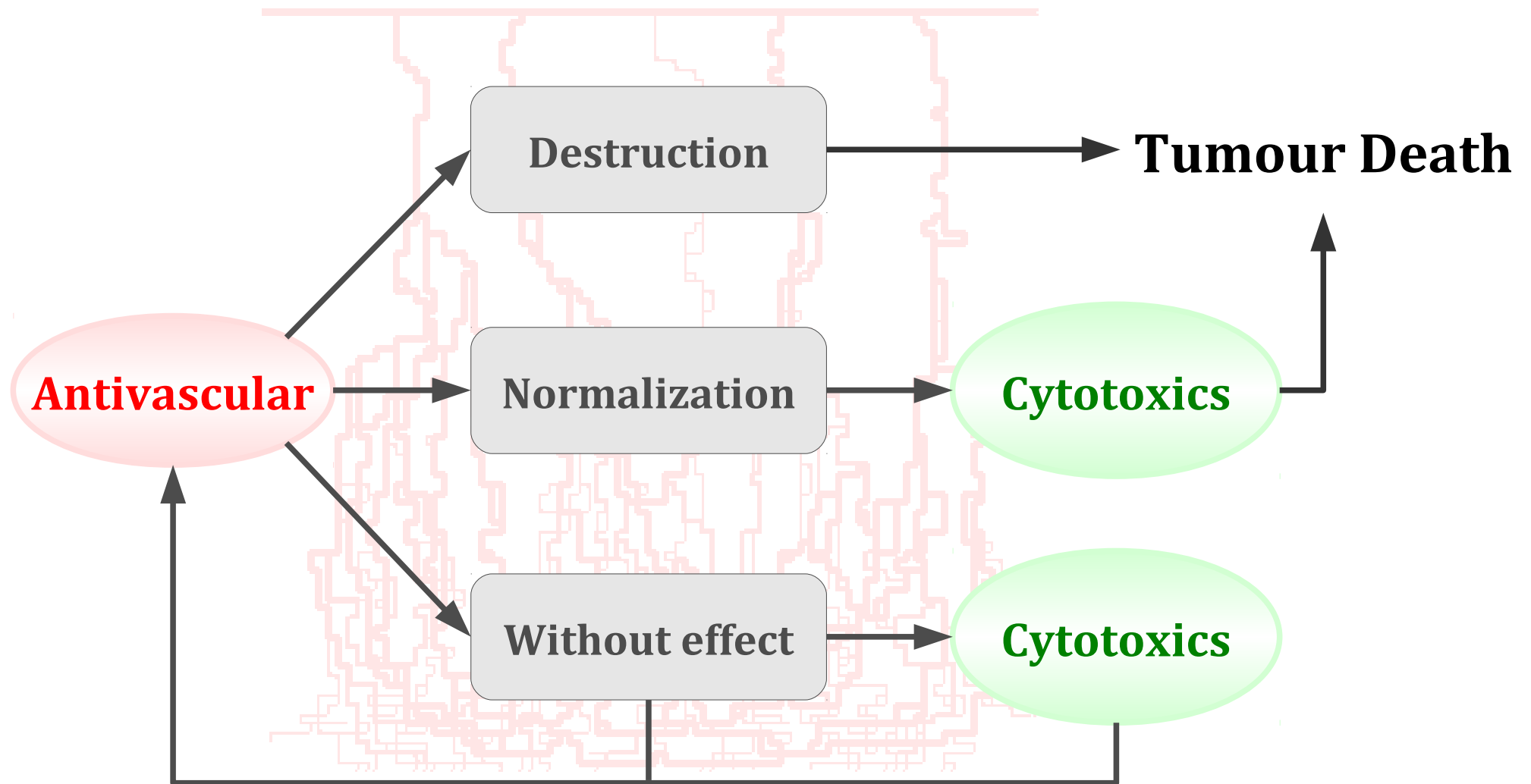


## Tumour case 2 (weakly vascularized)



# Therapeutics coupling

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# The next step ...

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## Phase 1

## Phase 2

### Experimental model

Sub-cutaneous implantation of U87-GFP tumours cells in matrigel

*Nude mice (immuno-deficient)*

Intra-cerebral implantation of C6 or 9L tumour cells

*Wistar rat*



### Virtual model

2D isotropic tissue

3D anisotropic brain tissue

### Control

Intravital fluorescence microscopy, immuno-histology

MRI, immuno-histology

### Action

Cytotoxics and antivascular

Cytotoxics, antivascular and radiotherapy



# Partners

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## Techniques for biomedical engineering and complexity

*Cell & tissue dynamics and functional microscopy*

Nicolas Glade, Arnaud Chauvière, Anne-Cécile Lesart (computational modelling)  
Marie-Paule Montmasson, Malika Hamel (experimental cell models, histology)  
Arnold Fertin, Yves Usson (image analysis)



## Clinatec *Biomedical research centre*

Flavien Caraguel , Boudewijn van der Sanden (*in vivo* models, intravital imaging)



## Grenoble Institute of Neurosciences

Emmanuel Barbier, Benjamin Lemasson (brain tumour models, MRI)  
François Estève (brain tumours, radiotherapy)



## Grenoble Images, Speech Signal and Control

Mazen Alamir, Mirko Fiacchini (control theory and optimization)



## Ecrins Therapeutics

Andrei Popov, Aurélie Juhem (development of antivascular molecules)

# Abstract

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The design of a patient-specific virtual tumour is an important step towards personalized medicine since the virtual tumour can be used to define the most adapted and efficient treatment protocol. However this requires to capture the description of many key events of tumour development, including angiogenesis, matrix remodelling, hypoxia, cell heterogeneity that will all influence the tumour growth kinetics and degree of tumour invasiveness. To that end, an integrated hybrid and multiscale approach has been developed based on data acquired on a preclinical mouse model as a proof of concept. Fluorescence imaging is exploited to build case-specific virtual tumours and to validate their spatiotemporal evolution. The validity of the model will be discussed as well as its potential to identify the best therapeutic strategy for each individual tumour case.